

Evaluation of muscle disorders after patients have been placed in a phenotypic pattern

Todd D. Levine¹, Richard J. Barohn², David S. Saperstein³, Jonathan S. Katz⁴, Mamatha Pasnoor⁵, Mazen M. Dimachkie⁵

¹HonorHealth Neurology - Bob Bové Neuroscience Institute

²University of Missouri School of Medicine, Columbia Missouri

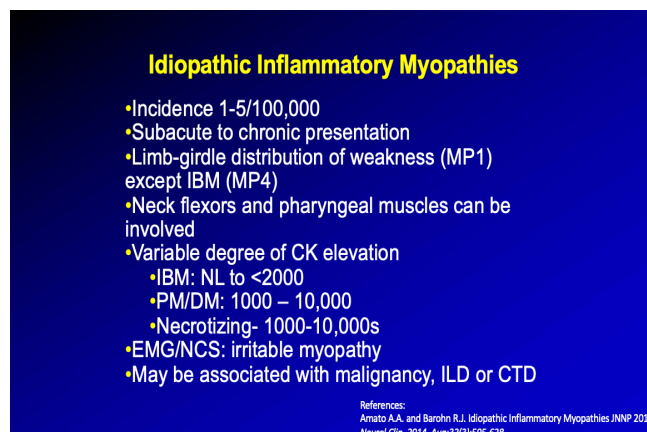
³Center for Complex Neurology, EDS and POTS Phoenix Arizona

⁴California Pacific Medical Center

⁵University of Kansas School of Medicine, Kansas City Kansas

Previously we discussed the pattern recognition approach to myopathy and neuromuscular junction disorders (see Pattern Recognition Approach to Neuromuscular Disorders: Myopathy & Neuromuscular Junction Disorders in the prior issue of the Barohn RRNMF Journal Volume 5 Issue 2 September 2024). That discussion was geared to teach how to approach the patient before ordering laboratory tests and putting patients into one or more of the ten phenotypic clinical patterns. The patterns are based solely on the history and presentation of the patient. In this lecture, a patient has already been placed into one or more of the possible myopathic patterns, specific muscle diseases should be considered as the next step. Therefore, it is time to think about ordering laboratory tests to support or at times confirm the clinical suspicion based on the pattern recognition. We will briefly review a number of myopathies, describe what are the most likely patterns with the condition, and discuss the appropriate laboratory studies, and the possible treatments. The evaluation and approach to neuromuscular junction disorders will be provided in a different lecture.

Inflammatory Myopathies Figure 1



From a historical perspective, the modern concept of inflammatory myopathies was introduced by Dr. John Walton in a 1954 article in *Brain* and the book titled *Polymyositis* authored by him and Dr. Raymond Adams in 1958. In the modern era, a classification of the inflammatory myopathies has been defined in the International Myositis Classification Criteria Project leading to the publication of the criteria by American College of Rheumatology / European Alliance of Associations for Rheumatology.

Idiopathic inflammatory myopathies are relatively rare, about 1 to 5 per 100,000. In most of them, the onset was sub-acute over months. Usually, they present for medical care in less than a year from the onset of symptoms except for inclusion body myositis (IBM) which has a delayed presentation and insidious onset over many years.

The typical pattern is limb-girdle (MP1 pattern). Because MP1 is the most common pattern, in some regards it is the least helpful because it does not distinguish between many different forms of muscle disease. An exception is IBM which presents with the MP4 pattern showing weakness in distal arms and proximal legs.

Figure 2

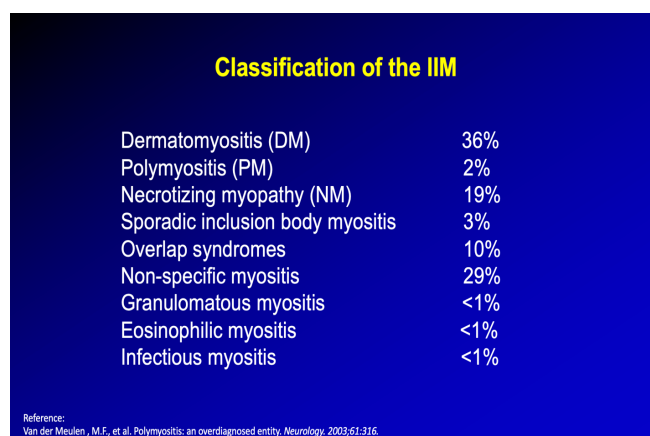
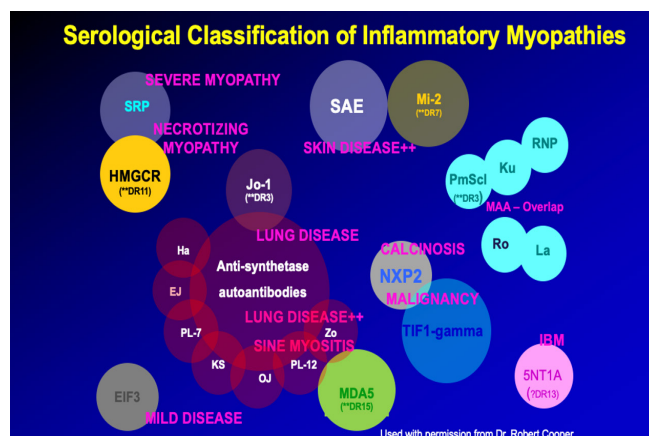


Figure 2 lists a number of different inflammatory myopathies and their frequencies according to a Dutch study. A common misbelief was that polymyositis is a frequent form of idiopathic inflammatory myopathy. But this is not the case. The Dutch study by Van der Meulen et al. showed that polymyositis accounted for only 2% of biopsy proved inflammatory myopathies. The most common inflammatory myopathy diagnosis in adults was dermatomyositis, followed by necrotizing myositis. Surprisingly, in this series, IBM made up only 3%. In our clinics in North America, IBM is more common than that. In the Database Evaluation for Muscle and Nerve Diseases (DEMAND) experience from four clinics, out of 490 inflammatory myopathy patients, 200 were IBM, 188 were polymyositis, and 102 were dermatomyositis.

Regarding laboratory studies, creatine kinase (CK) is very useful and its elevation indicates muscle damage. The magnitude of CK elevation can sometimes provide a clue as to the type of inflammatory myopathy. For example, IBM tends to have CK levels of 500 to 1000 IU/L range, whereas in polymyositis and dermatomyositis it is often over a thousand, and in necrotizing myopathy, it can be much higher. CK may be normal in DM and IBM.

Electromyography (EMG) is also helpful because it can reveal short-duration and small-amplitude motor unit potentials with irritability (denervation potentials in the form of fibrillations or positive sharp wave discharges). The presence of denervation potential suggests an active disease process. But their presence does not distinguish one muscle disease from another.

Figure 3



Some inflammatory myopathies are associated with specific serologic abnormal antibodies (Figure 3).

One of the first serologic antibodies that were discovered in inflammatory myopathy patients was the Jo-1 antibody. Subsequently, other tRNA synthetase antibodies were discovered. What is important about the tRNA-synthetase autoantibodies is that they predict a very high likelihood that the patient has concomitant interstitial pulmonary fibrosis. Both the muscle and the lung are

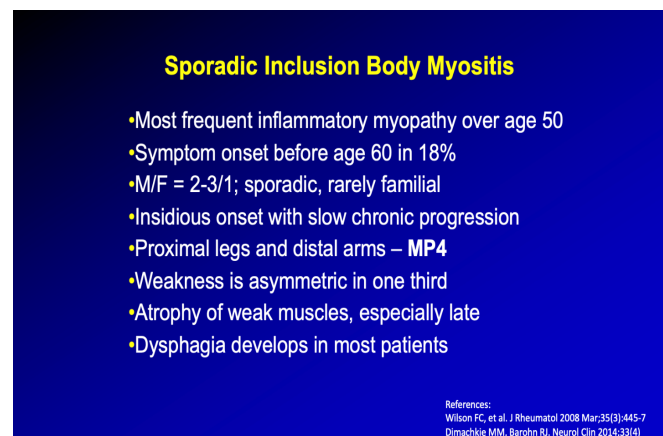
targets of the autoimmune attack. The autoimmune attack on the lungs tends to be difficult to reverse with treatment. Therefore, it is important to identify these patients early and treat them aggressively.

Some of these myopathies can be associated with an increased risk for malignancy up to 4 years from disease onset. While malignancies are overall more frequent in dermatomyositis, 2 myositis specific antibodies, Anti-NXP-2 (NXP2) and TIF1-gamma, are predictive of a higher likelihood of having an underlying malignancy. In dermatomyositis, SAE and Mi-2 antibodies are more commonly seen in patients with skin diseases such as calcinosis, severe alopecia, or other forms of breakdown of the skin.

Necrotizing myopathy is associated with SRP antibodies as well as HMGCR antibodies if the patient has been on a statin-lowering agent and sometimes without identifiable statin exposure. Finally, IBM is associated with NT5C1A antibodies in approximately half of the cases (See Figure 5 below and Barohn, Dimachkie, Jackson Neurol Clin 2014 and Dimachkie, Barohn, Amato Neurol Clin 2014).

Inclusion Body Myositis

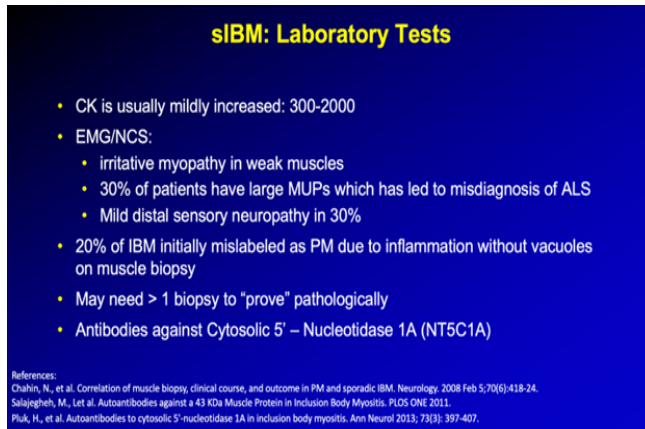
Figure 4



IBM in most series (other than the Dutch study noted above) is the commonest inflammatory myopathy in older adults. It is uncommon to occur before age 50. It is more common in men. IBM has a very slow chronic progression, and the average time between onset of weakness and diagnosis is approximately 8 years. These patients may not realize they have a muscle disease for many years. They may think their weakness is simply age-related because it comes on so slowly. IBM has a unique pattern of weakness involving the proximal legs and the distal arms. It is often asymmetric. The muscle involvement is very selective in the proximal legs, the quadriceps muscles (knee extensors) are predominantly involved whereas the hamstrings (knee flexors) are relatively spared. In the arms, the involvement is selective to muscles in the flexor compartment and

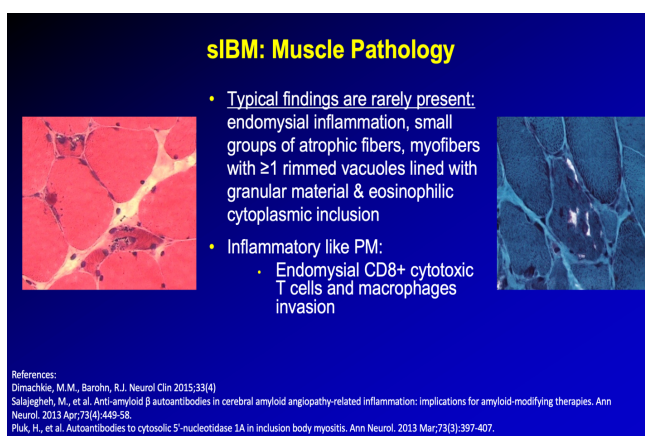
tends to spare the extensor compartment. Therefore, the weakness in the upper extremities involves finger flexors and wrist flexors. Some IBM patients also have dysphagia.

Figure 5



The CK is usually mildly elevated or normal in IBM. Many of these patients will have concomitant neuropathy based on symptoms and signs and electromyographic testing. The needle electromyography (EMG) findings can show long-duration and high-amplitude motor unit potentials which can make the diagnosis confusing and can lead to the misdiagnosis of amyotrophic lateral sclerosis (ALS). However, the pattern of weakness in IBM should make it very clear that it is unlikely to be ALS.

Figure 6

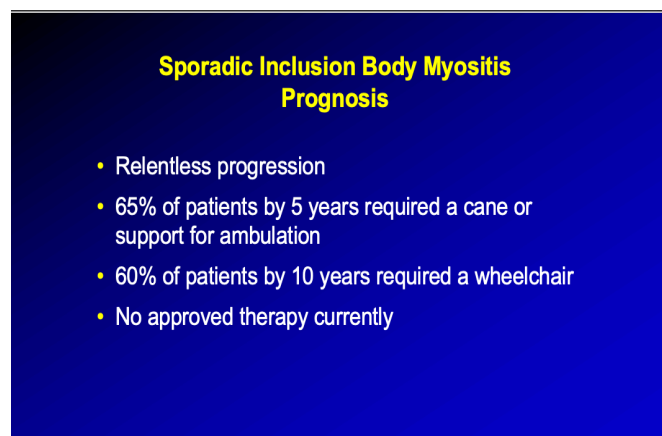


Historically, the standard way to diagnose IBM is via a muscle biopsy. Figure 6 shows the hematoxylin and eosin stain of a biopsied muscle from an IBM patient. There are small atrophic fibers that contain inclusions and small vacuoles. The vacuoles are easier to visualize on the trichrome stain. This type of biopsy finding is characteristic of IBM. However, sometimes the biopsy shows mild inflammation without the characteristic vacuoles. We believe the inflammation is likely a secondary phenomenon

as a result of degenerative processes ongoing within muscle fibers. This can therefore lead to the erroneous diagnosis of polymyositis if vacuoles and inclusions are not seen. If the patient is mistakenly diagnosed as polymyositis this can lead to them being put on corticosteroids and other immunosuppressant agents with no clinical benefit. Ultimately, the patient may undergo a second muscle biopsy to search for the characteristic vacuoles leading to the correct diagnosis of IBM. There is a serologic test, the NT5C1A antibody that can be positive in up to 50% of the patients with IBM. However other diseases can be associated with the NT5C1A. For example, patients with Sjogren's syndrome, systemic lupus erythematosus and even some neuromuscular disorders that are non-immune mediated may have a positive NT5C1A antibody titer. Therefore, in the right clinical setting where the patient has asymmetrical proximal leg and distal arm weakness with a slow progression, obtaining a positive serologic antibody test confirms the clinical suspicion and you can probably avoid a muscle biopsy.

This antibody test is now commercially available and should be obtained in any patient where there is a clinical suspicion of IBM based on the pattern recognition. However, the absence of the NT5C1A antibody does not rule out IBM and in this setting a muscle biopsy is necessary to confirm the diagnosis.

Figure 7



The course of IBM is very slow, relentless, and progressive. About 65% of patients need help with an assistive device for walking after five years and about 60% are in a wheelchair after 10 years.

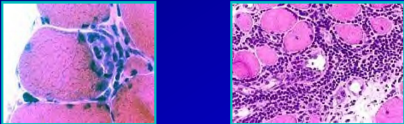
Unfortunately, there is no effective therapy. A recent phase 2/3 randomized control trial of the drug arimocloamol, a heat shock protein inducer, was negative (See Machado PM, et al. *Lancet Neurology* 2023;10:900-911).

Polymyositis

Figure 8

PM - Laboratory Features

- Serum CK usually elevated; 1000-20,000
- May be associated with myositis specific autoantibodies
- EMG/NCS: irritative myopathy
- Muscle biopsy:
 - Endomysial inflammation
 - Surround & commonly (63%) invade non-necrotic fibers expressing MHC antigens
 - Necrosis, phagocytosis & regenerating myofibers



Reference:
Dimachkie, M.M., Barohn, R.J., Amato, A.A. Idiopathic inflammatory myopathies. Neurol Clin. 2014 Aug;32(3):595-628, vii.

Polymyositis presents with the classic hallmark MP1 pattern of limb-girdle weakness as previously noted. In addition to limb-girdle weakness, there can be neck flexor weakness and dysphagia. Usually, the facial muscles are not involved. The pathology of polymyositis is different from IBM. There are many inflammatory cells surrounding nonnecrotic muscle fibers that otherwise appear to be healthy (Figure 8).

What used to be called polymyositis has splintered into 4 clinically, pathologically and serologically distinct diseases: necrotizing autoimmune myopathy, overlap syndrome, anti-synthetase syndrome and inclusion body myositis.

Figure 9

Myositis Specific Autoantibodies (MSA)

Autoantibody	Antigen	Antigen Function	Clinical Syndrome
Jo-1 Muscle in 90%	Histidyl tRNA	Protein Synthesis	ILD (50-75%) Mechanics hands Raynaud's, joint
PL-7	Threonine tRNA	Protein Synthesis	ILD (90%) GI (15%)
PL-12 Muscle in 52%	Alanyl tRNA	Protein Synthesis	ILD (90%) GI (20%)

We now define patients with Polymyositis and Anti-synthetase antibodies as having "An Anti-synthetase Syndrome"

Reference:
Dimachkie, M.M., Barohn, R.J., Amato, A.A. Idiopathic inflammatory myopathies. Neurol Clin. 2014 Aug;32(3):595-628, vii.

We believe this inflammatory cell invasion of non-necrotic fibers represents a cellular-mediated attack on the muscle fibers.

Some cases of polymyositis are associated with tRNA synthetase antibodies (Figure 9) suggestive of the anti-synthetase syndrome. Polymyositis with these antibodies is now known as anti-synthetase syndrome and is associated with interstitial lung disease. These patients need to be treated aggressively.

Necrotizing Myopathy

Figure 10

Necrotizing Myopathy

- Can be immune mediated or toxic
- Women/men = 3/1, onset age 30+
- Severe rapid progressive proximal weakness – **MP1**
- Triggers: drugs (**statins**), checkpoint inhibitors, fibrates, zetia, cyclosporine, labetalol, EtOH, propofol)
- More resistant to treatment than PM or DM especially when triggered by cancer or drug-induced
- CLAM – cholesterol lowering agent myopathy (Ringel 1991)
 - Early toxic cases
- SANAM – statin associated necrotizing autoimmune myopathy
 - Continued weakness 2 months after stopping statins

Reference:
Dimachkie, M.M., Barohn, R.J., Amato, A.A. Idiopathic inflammatory myopathies. Neurol Clin. 2014 Aug;32(3):595-628, vii.

In the group of immune-mediated myopathies, necrotizing myopathy is the most recently identified. There are two forms of necrotizing myopathy. One is an acute toxic myopathy which is typically caused by drugs or toxins, and the other is a more chronic immune-mediated necrotizing myopathy. The most common class of drugs to cause acute necrotizing myopathy are the cholesterol-lowering agents. In fact, this was first identified in 1991 and was called cholesterol-lowering agent myopathy (CLAM). Rarely, acute statin induced myopathy can present as a rhabdomyolysis with myoglobinuria (MP8), but more often as a subacute MP1 pattern without rhabdomyolysis (See Barohn, Dimachkie, Jackson Neurol Clin 2014). Usually, once the offending toxic drug is withdrawn the toxic myopathy resolves. However, some patients continued to have progressive myopathy symptoms and signs and persistently elevated CK for 1-2 months after the cholesterol-lowering agent was stopped. We call these cases statin-associated necrotizing autoimmune myopathy

(SANAM) and it is believed that the statin drugs set off an immune-mediated process against muscle fibers. Other drugs or agents that can cause acute toxic myopathy include alcohol, cyclosporine, propofol or immune checkpoint inhibitors (see Figure 8 in Barohn, et al, The Pattern Recognition Approach to Neuromuscular Disorders: Volume 5 Issue 2 September 2024).

Figure 11

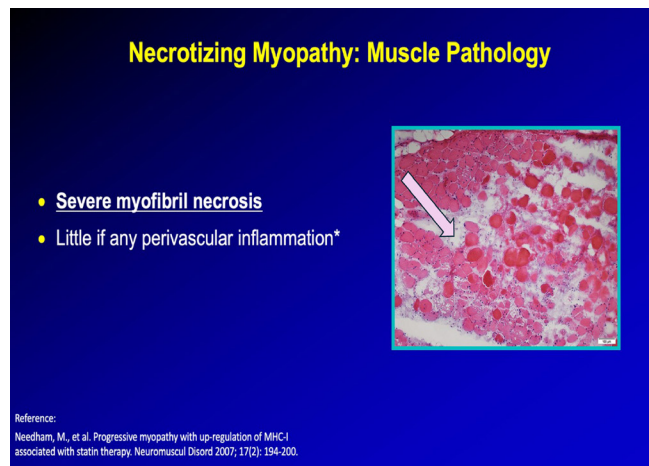


Figure 11 shows the pathology of a severe necrotizing myopathy. There is little inflammation. There is a great deal of muscle fiber necrosis. There are “ghost fibers” that have replaced healthy muscle fibers (See arrow in Figure 11).

Figure 12

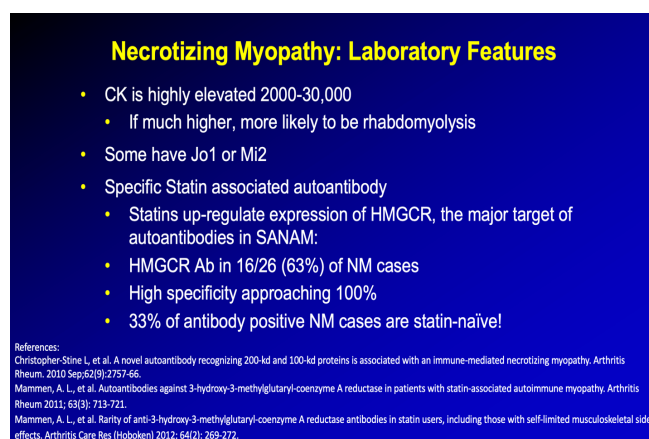


Figure 13

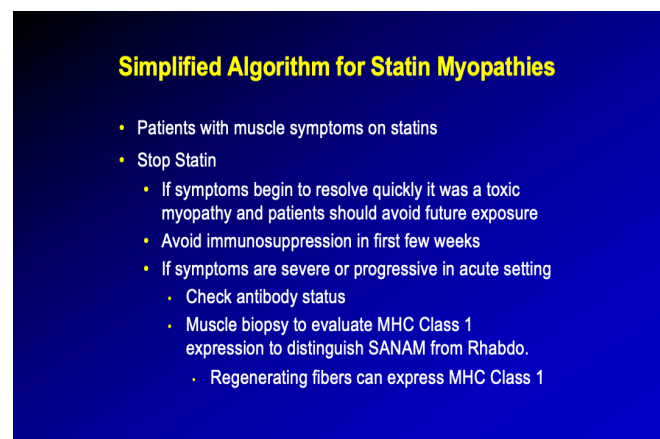
Necrotizing Autoimmune Myopathy Specific Autoantibodies			
Autoantibody	Antigen	Antigen Function	Clinical Syndrome
SRP 16% of NAM	SRP RNA complex	Protein translocation	Acute and severe NM in the fall season, difficult to treat,
HMGCR 40% of NAM	Reductase	Cholesterol biosynthesis	Immune NM with or without obvious statin use

In necrotizing myopathy, CK level tends to be high, at least 1,000 IU/L sometimes over 10,000 IU/L (Figure 12).

HMGCR autoantibody is present in about two thirds of SANAM (Figures 12 and 13). It is speculated that the statins upregulate the expression of HMGCR. However, it has also been demonstrated that about one third of patients with necrotizing myopathy and HMGCR antibodies have never been previously exposed to a statin medication. So clearly in some cases, the statin alone is not the precipitating event for the autoimmune process.

Another antibody associated with necrotizing myopathy is SRP antibodies. These cases tend to be seasonal, often following a flu-like illness.

Figure 14




How should you handle a patient who is on statin and has muscle weakness? (Figure 14)

If the patient on a statin presents with muscle cramps and muscle aches without objective weakness the first thing to do is to stop the statin. We would recommend obtaining a serum CK to determine if there is evidence of muscle fiber damage. Usually, these patients present within eight weeks of starting the statin. If the statin is stopped and they improve there is nothing further to do other than get an additional CK if the initial one was elevated. If the statin is stopped and they do not start to get better after 8 weeks, we recommend to test for HMGR antibody and trend CK levels. If the course continues to worsen, there is a need to consider a muscle biopsy. If the HMGR antibody is elevated an argument can be made not to perform a muscle biopsy. In either case, immunosuppressive therapy for SANAM should be initiated.

Dermatomyositis Figure 15

Diagnosis of Dermatomyositis

- Proximal muscle weakness (MP1)
- CK can be normal (amyopathic) or as high as 10,000
- Irritative myopathy by EMG
- Myositis specific autoantibodies in 2/3 of patients
- Skin changes



Reference:
Dimachkie M.M., Barohn R.J., Amato A.A. Idiopathic inflammatory myopathies. Neurol Clin. 2014 Aug;32(3):595-628, vii.

Dermatomyositis has an MP1 pattern of presentation. CK is generally very high but cases with a normal CK and typical skin involvement do occur and have been called amyopathic dermatomyositis. The red skin discoloration usually appears on the extensor surfaces of the fingers, elbows or arms, as well as on the front and back of the neck (Figure 15). The facial rash can appear on the forehead, cheeks, and eyelids. EMG in dermatomyositis shows an irritable myopathy.

Figure 16

Dermatomyositis

- Complement-mediated autoimmune microangiopathy
- Affects both children and adults
- Subacute or chronic presentation
- May be associated with underlying malignancy
- Limb-girdle distribution of weakness with:
 - Erythema, scaling rash over malar area of face, extensor joints, MCP and IP joints (Gottron's papules)
 - Heliotrope rash, periorbital edema
 - Joint contractures
 - In children: calcinosis, vasculitis

Reference:
Dimachkie M.M., Barohn R.J., Amato A.A. Idiopathic inflammatory myopathies. Neurol Clin. 2014 Aug;32(3):595-628, vii.

Unlike polymyositis, dermatomyositis is an autoimmune muscle disease that can occur in children (Figure 16). Other features of dermatomyositis include calcinosis, vasculitis, and joint contractures.


Before the 1950s and the availability of corticosteroids, children who had dermatomyositis tended to die very often due to ischemic bowel disease because of vasculitis and its widespread nature. Therefore, juvenile dermatomyositis is vasculitis of the muscle rather than myositis. B lymphocytes invade and circle the small capillaries inside of the muscle fascicles. There is immunoglobulin (IgM) and complement deposition around the blood vessels. This leads to entire muscle fascicles that become ischemic.

The blood vessels course through the middle of the fascicle and therefore muscle fibers on the edge of a muscle fascicle become ischemic and become smaller. That leads to the appearance perifascicular atrophy (Figure 17).

Figure 17

Dermatomyositis: Muscle Biopsy

Perifascicular atrophy



B cell predominance in perivascular infiltrates perivascular deposits of IgM and complement

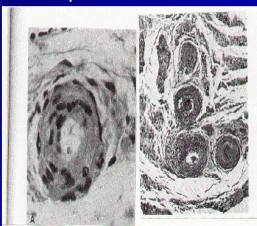


Figure 18

DM Specific Autoantibodies			
Autoantibody	Antigen	Antigen Function	Clinical Syndrome
Mt-2	Helicase	Nuclear transcription	Nail fold lesions; higher in Hispanics
Anti-p155(140) TIF1 γ	Transcription intermediary factor 1 γ	Part of tripartite-motif (TRIM) family; interacts with SMAD 2,3,4; muscle cell regeneration	Cancer in adults; Severe skin disease in children; palmar hyperkeratotic papules, psoriasis like
MDA-5 aka CADM-140 May be weak!!	Type I IF-inducible protein 1, IFIH1	Positive regulator of the IFN response	Severe ILD & cardiopulmonary syndrome, skin ulceration, tender palmar papules
MJ (p140) Up to 40% of DM	NXP-2	Nuclear transcription	JDM with calcinosis
Anti-small ubiquitin-like modifier 1	Small ubiquitin-like modifier 1	post-translational modification; not targeting proteins for degradation	Skin presenting before muscle manifestations; dysphagia common

Figure 18 shows a number of autoantibodies that have been identified in a number of cases of dermatomyositis. Some of these cases predict interstitial lung disease and others can predict severe skin disease.

Cancer and Inflammatory Myopathies

Since many inflammatory myopathies (including IBM) are associated with an increased risk for cancer and are paraneoplastic (Figure 19), what is the standard cancer screening in a patient with inflammatory myopathy? (Figure 20)

We recommend that cancer surveillance screening be performed yearly for the first 5 years after diagnosis of inflammatory myopathy. There are two approaches of cancer surveillance as outlined in Figure 20 or a combination of these approaches can be used.

Figure 19

Cancer Screening in PM/DM/NM	
<ul style="list-style-type: none"> Risk is highest within the first year of diagnosis In those with polymyositis, the risk fell to expected rates 5 years after diagnosis In those with dermatomyositis, the risk is always elevated <ul style="list-style-type: none"> Risk of ovarian, pancreatic, lung cancer remained elevated for up to 5 years Pancreatic and colorectal cancer risks remained elevated past 5 years 	<p>Reference: Hill, C.L. et al. Frequency of specific cancer types in dermatomyositis and polymyositis: a population-based study. <i>Lancet</i>. 2001 Jan 13;357(9250):96-100.</p>

Figure 20

Cancer Screening in DM/PM/NM	
<ul style="list-style-type: none"> A matter of debate Best practice guidelines say to repeat yearly for 1st 5 years after diagnosis Approach #1 <ul style="list-style-type: none"> Complete history and physical exam Routine blood and urine tests Fecal occult blood Chest x-ray; mammogram Additional tests in case of specific signs and symptoms 	<p>Approach #2: WHOLE BODY CT OR PET</p>

Figure 21 outlines the various immunosuppressive drug therapy options for autoimmune inflammatory myopathy. For polymyositis, dermatomyositis and autoimmune necrotizing myopathy, corticosteroids and intravenous immunoglobulin (IVIG) are often used as first-line immunotherapies. There is now an FDA-approved indication for IVIG in dermatomyositis.

Figure 21

PM/DM/NM Drug Therapy	
<ul style="list-style-type: none"> 1st Line <ul style="list-style-type: none"> Prednisone IV methylprednisolone IVIG* 2nd Line <ul style="list-style-type: none"> Methotrexate Azathioprine* Mycophenolate mofetil 	<ul style="list-style-type: none"> 3rd Line <ul style="list-style-type: none"> Rituximab* (Oddis) Cyclophosphamide Etanercept* (Amato) Tacrolimus (Oddis) Cyclosporine <p>*RCT</p> <p>References: van der Ploeg, et al. Rituximab therapy for myopathy associated with anti-signal recognition particle antibodies: a case series. <i>Arthritis Care Res (Hoboken)</i> 2010; 62(9): 1328-1334. Amato, A.A., et al. A randomized, pilot trial of etanercept in dermatomyositis. <i>Ann Neurol</i> 2011; 70(3): 427-436. Dimachkie, M. M., Barohn, R. J., Amato, A. A. Idiopathic inflammatory myopathies. <i>Neurol Clin</i> 2014; 32(3): 595-628, vii. Baraliakos, X., et al. Efficacy and safety of upadacitinib in patients with active psoriatic arthritis and axial involvement: results from two phase 3 studies. <i>Arthritis Res Ther</i>. 2023 Apr 10;25(1):56.</p>

Pompe's Disease

Figure 22

Pompe Disease

- Autosomal recessive disorder caused by deficiency of lysosomal alpha-glucosidase, also called acid maltase deficiency
- Infantile
 - Cardiac symptoms, hypotonia, hepatomegaly, macroglossia, failure to thrive
 - Fatal by 2 years of age
- Juvenile onset
 - Symptoms before 10 years of age
 - Limb girdle weakness, waddling gait (MP1), respiratory weakness (MP7) with death by 30 years of age
- Adult Onset
 - Symptoms age 18-65, proximal muscle weakness (MP1); scapuloperoneal (MP3)
 - Can be confused for PM or LGMD

**** Respiratory weakness can be initial or predominant symptom due to selective diaphragm involvement (MP7)

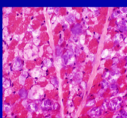
Reference:
Dasouki, M., et al. Pompe disease: literature review and case series.
Neurol Clin 2014; 32(3): 751-776, ix.

Pompe disease is also known as alpha-glucosidase deficiency or acid maltase deficiency. Pompe disease is an autosomal recessive disorder. It can present at various ages. If it presents in infancy the disease used to be often fatal by 2 years of age without enzyme replacement therapy. Juvenile onset patients used to die of the disease by the second or third decade without enzyme replacement therapy. The late-onset Pompe disease with an adult presentation usually has an MP1 pattern however can also present with an MP3 scapuloperoneal pattern. Because Pompe disease most often has an MP1 presentation it can mistakenly be diagnosed as polymyositis. As also pointed out in the pattern recognition review, Pompe disease patients often have prominent respiratory issues due to diaphragm involvement (MP7).

Figure 23

Pompe Disease

- Laboratory
 - Moderate elevation in CPK, but may be normal
 - EMG with increased insertional activity.
 - ****Myotonia on EMG should raise suspicion
 - Decreased alpha-glucosidase from dried blood spot is recommended test.
 - Specific genetic testing if abnormal
 - Muscle biopsies have characteristic vacuolar appearance and PAS shows large deposits of glycogen



Reference:
Dasouki, M., et al. Pompe disease: literature review and case series.
Neurol Clin 2014; 32(3): 751-776.

The serum CK is usually mild to moderately elevated. The EMG is interesting because in addition to short-duration small-amplitude motor unit potentials and fibrillation potentials, there can be the presence of myotonic potentials. Figure 23 shows a muscle biopsy showing H&E stained section with vacuolated muscle fibers and loss of muscle fibers and large deposits of glycogen accumulation resulting from a lack of breakdown due to enzyme deficiency.

Figure 24

Pompe Disease

- Alpha-glucosidase approved in 2006 for childhood onset.
- Approved in 2010 for Adults
- Prevent further loss of muscle function
 - Improved six-minute walk test as well as delays progression of loss in FVC
- Bi-weekly infusions
- Monitor for IgG antibodies every 3 months for years and then annually after that

References:
Davison, M., et al. Recombinant human acid [alpha]-glucosidase: major clinical benefits in infantile-onset Pompe disease. Neurology 2007; 68(2): 99-109.
Morgan, C., et al. A randomized study of alglucosidase alfa in late-onset Pompe's disease. N Engl J Med 2010; 362(15): 1395-1406.

One of the biggest breakthroughs in modern medicine is the introduction of enzyme replacement therapy for Pompe disease. This was the first FDA-approved molecular therapy for a muscle disease. Initially, it was demonstrated that enzyme replacement therapy could dramatically improve infants with Pompe disease and preserve life. It was subsequently shown that chronic enzyme replacement therapy can slow down the progression of juvenile and adult Pompe disease and improve muscle function. Therefore, it is important to recognize Pompe disease early. As the possibility of enzyme replacement therapy was being studied in clinical trials, measuring alpha-glucosidase activity from a phlebotomy sample was developed. This is now a standard procedure for all patients being suspected of Pompe disease. Further genetic testing confirms the diagnosis. Therefore, the combination of an abnormally low alpha-glucosidase level on a blood sample and an abnormal genetic test for Pompe disease can avoid a muscle biopsy and enzyme replacement therapy can be initiated.

Channelopathies

Figure 25

Channelopathies

- Caused by mutations in ion channels: chloride, sodium, calcium or potassium
- Inherited as sporadic or autosomal dominant disorders
- Can cause increased or decreased excitability of sarcolemma
- Divided into nondystrophic myotonias (chloride or sodium channels) or periodic paralyses (sodium or calcium channels, also potassium in ATS or TT-PP)
- Onset typically within first two decades of life
- Do not shorten life but cause significant effect on QOL
- Often misdiagnosed as functional etc.

References:
Trivedi, J. R., Barohn, R. J., Bundy, B., Statland, J., Salajegheh, M., Rayan, D. R., Venance, S. L., Wang, Y., Fialho, D., Matthews, E., Cleland, J., Gorham, N., Herbelin, L., Cannon, S., Amato, A., Griggs, R. C., Hanna, M. G. Non-dystrophic myotonia: prospective study of objective and patient reported outcomes. *Brain* 2013; 136 (Pt 7): 2189-2200.
Statland, J. M., et al. Mexiletine for symptoms and signs of myotonia in nondystrophic myotonia: a randomized controlled trial. *Jama* 2012; 308(13): 1357-1365.

Another group of muscle disorders are the disorders due to muscle channel dysfunction (Figure 25). This group of genetic disorders can be due to mutations in either the chloride, sodium, calcium, or potassium channels. They are inherited disorders that can either increase or decrease the excitability of the muscle fiber membrane. The two most prominent presentations of channelopathies are episodic weakness after exercise or at times unrelated to exercise (MP9), or muscle stiffness and decreased ability to relax (MP10).

Figure 26

Periodic Paralysis

<u>Hyperkalemic</u>	<u>Hypokalemic</u>
<ul style="list-style-type: none"> • Mutations in SCN4A • Attacks last minutes to hours • Precipitated by fasting, rest after exercise, or eating high potassium foods • Potassium greater than 5 during attacks often but not always present • During attacks patients are areflexic, no effect on cardiac or respiratory muscles • Rx for prevention of attacks <ul style="list-style-type: none"> • dichlorphenamide now FDA approved • acetazolamide 	<ul style="list-style-type: none"> • Mutations in CACNA1S or SCN4A or KCNJ2 (ATS) • Attacks can last for hours to days • Triggers: alcohol, carbohydrate rich foods, stress, rest after exercise • Potassium during attacks less than 3.0 • Can develop fixed muscle weakness late in life • Treatment during attacks is to give potassium • Prevention: consume low carbohydrate diets • Rx for prevention of attacks <ul style="list-style-type: none"> • dichlorphenamide now FDA approved; • acetazolamide

We divide the muscle channelopathies into two groups. One group is the nondystrophic myotonias due to chloride and sodium channel mutations. The other group is the periodic paralysis. These conditions often begin before the first two decades of life and they generally do not cause permanent weakness which is why they are not considered dystrophic. There are exceptions however, as some middle and late age patients with long-standing episodic symptoms of periodic weakness can eventually develop permanent weakness. These conditions do affect the patient's quality of life. These conditions can be difficult to diagnose and

sometimes patients can be labeled as being functional or psychogenic.

Periodic paralysis can be further divided into two groups based on the serum level of potassium: hyperkalemic and hypokalemic (Figure 26). The hyperkalemic-associated episodes tend to last a shorter time than the hypokalemic subtype. Hyperkalemic paralysis is typically precipitated by fasting, resting after exercise, or eating high-potassium foods. Hypokalemic paralysis is typically precipitated by alcohol intake, carbohydrate-rich food, stress, and resting after exercise.

The evaluation of channelopathies is outlined in Figure 27.

Figure 27

Channelopathies: Evaluation

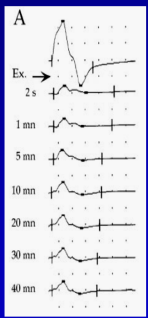
- Measure K⁺ during attack
- Episodic symptoms of myotonia or paralysis
 - EMG
 - May be normal
 - May show myopathic units
 - May show myotonia
- Short exercise test – myotonia
- Long exercise test – periodic paralysis
- No longer do K⁺ or insulin/glucose challenges
- Genetic studies
- R/O secondary forms

Examining a patient and measuring the potassium level during the paralysis attack is optimal but it is often difficult to have these opportunities. In hyperkalemic paralysis, EMG between attacks may be normal or may show myopathic units or myotonic discharges. However, these are not observed in the hypokalemic group. On nerve conduction studies, the long exercise test for periodic paralysis and the short exercise test for non-dystrophic myotonias can sometimes be useful to differentiate the subtypes (Figure 28).

Figure 28

Long Exercise Test for Periodic Paralysis

- Record ulnar CMAP Amp baseline
- Exercise ADM 5 min
- Check CMAP every 2 min. for 50 min
- In PP (all types), over next 10-40 min, grad dec amp



References:
McManis, P.G., et al. The exercise test in periodic paralysis. *Muscle Nerve*. 1986 Oct;9(8):704-10.
Fournier, E., et al. Electromyography guides toward subgroups of mutations in muscle channelopathies. *Ann Neurol*. 2004 Nov;56(5):650-61.

The short exercise test can sometimes be used to monitor electrophysiologic improvement after treatment initiation for myotonia (Figure 29).

Figure 29

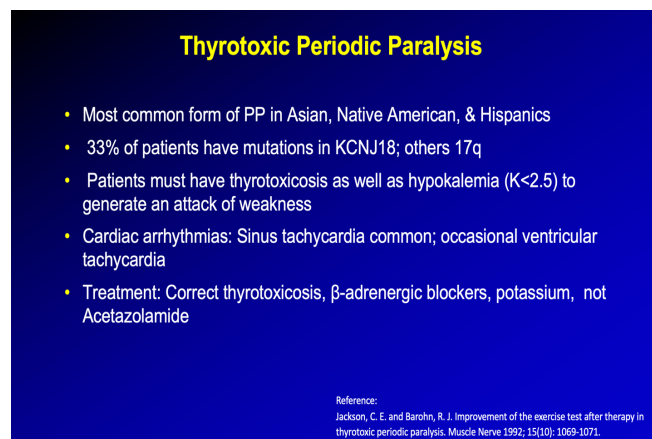


The most efficient and accurate way to diagnose channelopathies is via genetic testing for sodium, calcium, chloride, or potassium channel mutations which are now commercially available. Prior to the advent of genetic testing, an attack could be provoked by administering potassium to produce hyperkalemia and then administering insulin with glucose to produce hypokalemia. However, this is no longer needed with the availability of genetic testing.

Therapy for periodic paralysis is either dichlorphenamide which is now FDA-approved or acetazolamide. Therapy for myotonia is sodium channel-blocking agents. For decades the drug of choice was mexiletine, but more recently ranolazine and lamotrigine have been shown to be effective (See Vivekanandam, et al. Lancet Neurology 2024;10:1004-1012).

Thyrotoxic Periodic Paralysis

Figure 30



There are secondary forms of periodic paralysis. The most common etiology for secondary periodic paralysis is thyrotoxic periodic paralysis.

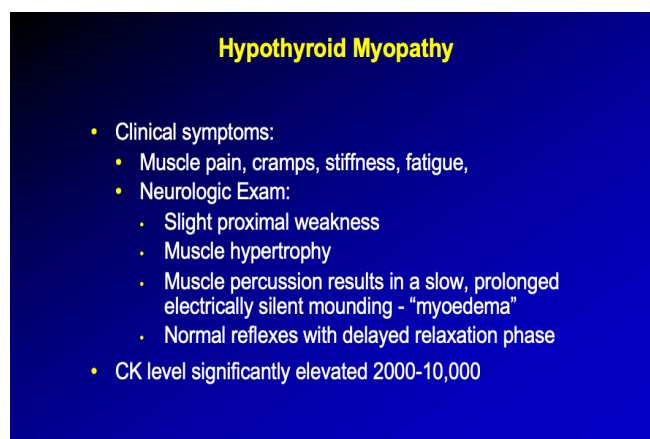
This is the most common form of paralysis in Asians, Native Americans, and Hispanics.

Many of these patients have a concomitant potassium channel defect on chromosome 18 or on chromosome 17. Clinically they have obvious thyrotoxicosis when they present with weakness and hypokalemia.

Acutely the treatment is to correct the hypokalemia, but the long-term treatment is to correct the thyrotoxicosis.

Hypothyroid Myopathy

Figure 31

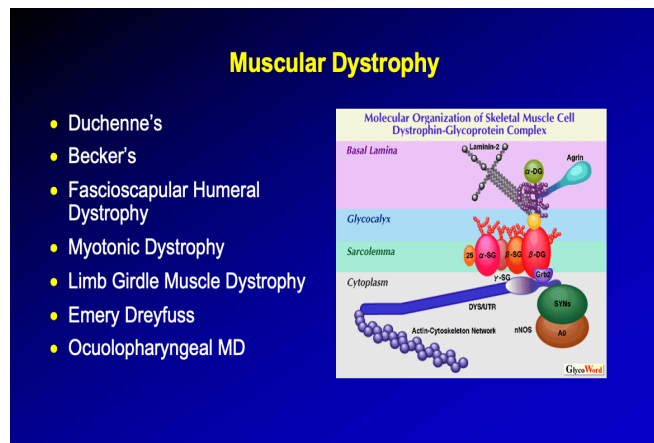


Hypothyroid myopathy patients present with cramps, stiffness, fatigue, and proximal weakness (MP1). Reflexes can exhibit a delayed relaxation phase. Serum CK is often very high. It is important to check thyroid functions in patients with unexplained elevated CK as some of these patients may not have a great deal of symptoms and signs of muscle disease or hypothyroidism.

Muscular Dystrophies

A number of different muscular dystrophies along with a diagram of the molecular organization of a skeletal muscle cell and the dystrophin glycoprotein complex is shown in Figure 32.

Figure 32

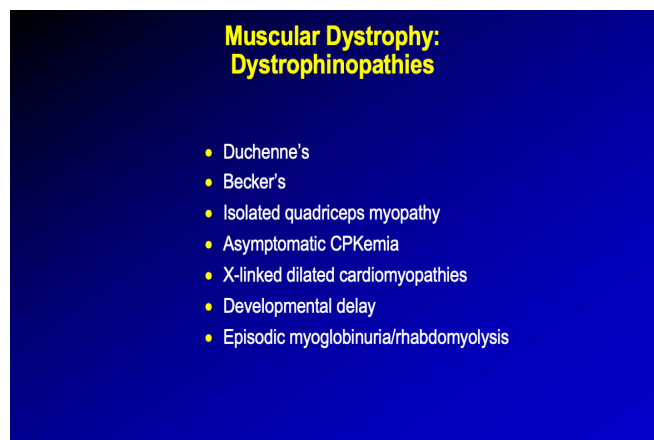


Dystrophinopathies (Duchenne's and Becker's Muscular Dystrophy)

Dystrophinopathies have a number of clinical presentations. The most common presentation is an MP1 pattern of muscle weakness for both Duchenne's and Becker's muscular dystrophy.

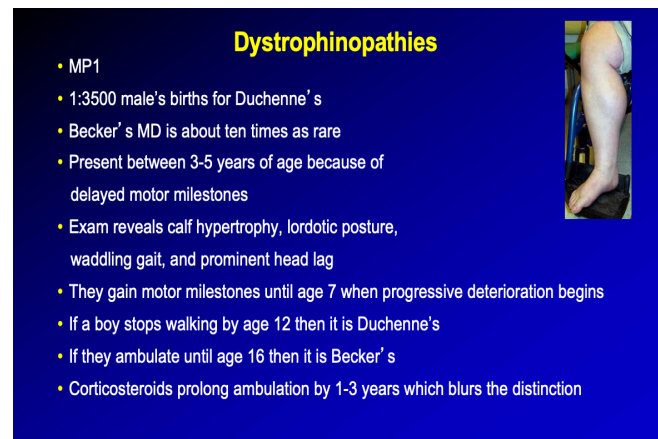
Duchenne and Becker's muscular dystrophy are due to genetic defects in the dystrophin gene and are therefore called dystrophinopathies. Other reported presentations of dystrophinopathies are shown in Figure 33.

Figure 33



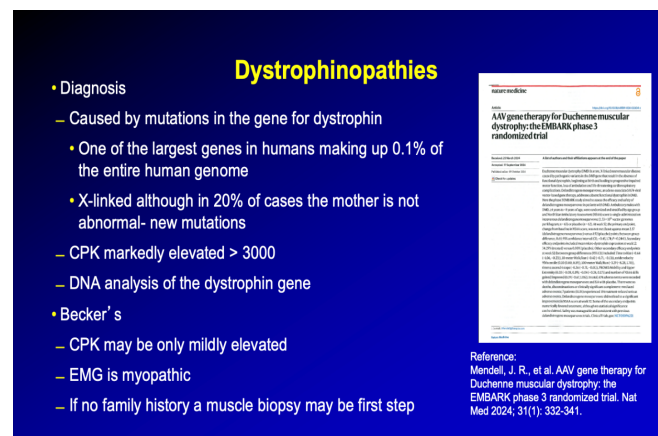
Typical patients with Duchenne muscular dystrophy present between ages 3 and 5 because of delayed motor milestones. There is often the presence of calf hypertrophy (Figure 34).

Figure 34



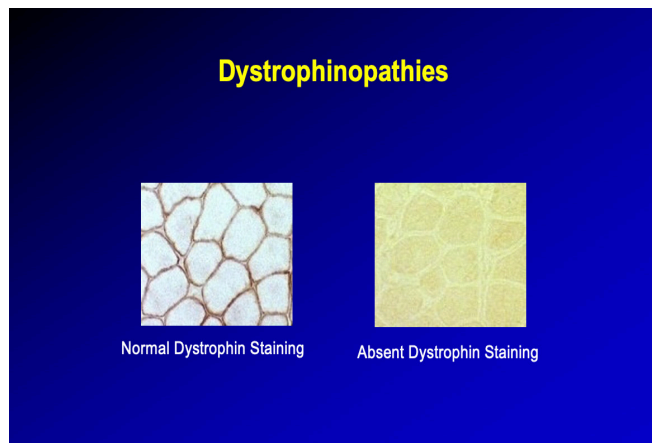
Boys usually stop walking by age 12 but the use of corticosteroids will prolong ambulation. There is now FDA-approved exon-skipping antisense therapy for dystrophinopathies due to a number of specific deletions in the dystrophin gene. More recently the FDA approved intravenous AAV mediated gene therapy (Elevidys). Therefore, accurate and early genetic diagnosis is important (Figure 35).

Figure 35



Muscle biopsies will show the absence of dystrophin, but these are no longer needed with a documented dystrophin mutation in the blood (Figure 36).

Figure 36



Facioscapulohumeral Dystrophy (FSHD)
Figure 37

Fascioscapulohumeral Dystrophy

- Focal weakness of the face, periscapular, biceps and triceps, and distal legs (tib ant or gastric); can be asymmetric (MP3)
- Spares deltoids despite pronounced scapular weakness
- MP3
- Often very asymmetric
- Incidence may be as high as 13:100,000
- Autosomal dominant but up to 25% of cases have no family history
- Onset in teens or early 20s
- CK levels can be normal up to 1000
- Genetic defect is complex but majority have decrease in tandem repeats in 4q35 - D4Z4 gene

Reference:
Statland, J. and Tawil, R. Facioscapulohumeral muscular dystrophy. Neurol Clin 2012; 32(3): 721-728, ix.

FSHD patients have a scapulooperoneal or MP3 weakness distribution (Figure 37). Patients have weakness of the face, periscapular muscles, biceps, and triceps. In these patients, the deltoid can appear normal with very atrophic biceps and triceps. Scapular winging is prominent and can be observed with either shoulder flexion or abduction. Most patients will provide an autosomal dominant family history but up to 25% will not. The diagnosis is based on demonstrating the genetic defect which is a decrease in the number of tandem repeats in the 4q35-D4Z4 gene.

Myotonic Dystrophy
Figure 38

Myotonic Dystrophy

- Most common muscular dystrophies in adults
 - Prevalence as high as 20:100,000
- DM1 is a multisystem disorder
 - Weakness, cataracts, myotonia before age 50
 - Weakness primarily affects face, oropharyngeal muscles, finger flexors, ankle/toe dorsiflexors sometimes quads & finger flex (like IBM)
 - Pronounced grip myotonia with a 1-5 second delay in relaxing
 - Multi-colored bright Christmas tree cataracts
 - Cardiac arrhythmias and conduction block lead to early mortality
 - Respiratory involvement
 - Cognitive involvement & brain MRI changes
 - Common to have endocrine involvement including diabetes, thyroid, testosterone deficiency, hyperlipidemia

Reference:
Thornton, C. A. Myotonic dystrophy. Neurol Clin 2014; 32(3): 705-719.

Myotonic dystrophy is probably the most common form of muscular dystrophy in adults. There are two forms of myotonic dystrophy. Myotonic dystrophy type 1 (DM1) is the classic form with multisystem involvement. Along with weakness, they have cataracts. They develop cardiac arrhythmias and conduction blocks which lead to very early mortality, often in the sixth decade. Some of these patients require pacemakers and automatic implantable cardioverter defibrillators. They can have various endocrine disorders, including diabetes, hypothyroidism, and testosterone deficiency. Cognitive impairment is very common.

Myotonic dystrophy can have a number of different presentation patterns including MP1, distal weakness MP2, ocular ptosis MP5, and muscle stiffness (MP10). Rarely, an adult DM1 patient can have severe knee extension weakness combined with severe finger flexor weakness MP4 pattern which is most often seen in IBM. However, the age of the patient and other typical myotonic dystrophy features help distinguish these two diseases.

Figure 39

Myotonic Dystrophy

- Myotonic Dystrophy Type 2 –DM2
 - Presents in middle age or later
 - Muscle pain and stiffness
 - Prominent proximal muscle weakness that often involves finger flexors
 - MP1, MP10
 - Spares facial muscles
 - EMG myotonia may not be present
 - Less common to experience the multisystem involvement

Myotonic dystrophy type 2 (DM2) presents later in life (Figure 39). They do not have all of the features typical for DM1. Most of these patients present with muscle pain and stiffness. On exam, they can have mild or prominent limb-girdle weakness (MP1).

One clue to DM2 is that very often their finger flexors are weak as well, but they do not have facial involvement and they may not have myotonia on exam.

Figure 40

Myotonic Dystrophies: Diagnosis

- CK in both may be mildly elevated
- Genetic testing confirms the diagnosis and obviates the need for muscle biopsy
- DM1 caused by CTG repeat in gene for dystrophin myotonia protein kinase (DMPK)
 - Number of repeats correlates inversely with age of onset and severity of disease
- DM2 caused by CCTG repeat in zinc finger protein 9 gene (ZNF9)
 - Number of repeats does not correlate with age of onset or severity
- In both disorders the aberrant RNA transcripts accumulate into nuclear aggregates
- Antisense oligonucleotide research studies in progress
- Mexiletine helps symptomatic myotonia

References:
 Thornton, C. A. Myotonic dystrophy. *Neural Clin* 2014; 32(3): 705-719, viii.
 Logigian, E. L., et al. Mexiletine is an effective antimyotonia treatment in myotonic dystrophy type 1. *Neurology* 2010; 74(18): 1441-1448.

The definitive diagnosis of both types of myotonic dystrophy is via genetic testing of the blood or muscle. Genetic testing confirms the diagnosis. DM1 is caused by CTG repeat expansion of the DMPK gene. DM2 is caused by CCTG repeat expansion of the ZNF9 gene (Figure 40).

In DM1 the number of repeats correlates inversely with the onset age and the severity of the disease. In DM2 the number of repeats does not correlate well with the onset of severity.

Both seem to relate to the increase in RNA transcripts that build up because of these extra repeats and there are several research studies now trying to decrease the amount of RNA with antisense nucleotides. EMG can show classic myotonia but is often more subtle in DM2 and CK may be mildly elevated.

Although not FDA-approved, mexiletine is very helpful to treat their symptoms of muscle stiffness and pain but it does not help weakness.

Oculopharyngeal Muscular Dystrophy

Figure 41

Oculopharyngeal Muscular Dystrophy

- Presents with ptosis, limited EOM and dysphagia in the 5th and 6th decade
 - MP5, MP7
- Can be mistaken for myasthenia gravis
- Mitochondrial disorders and myotonic dystrophy are also in the differential
- Often lead to multiple blepharoplasty procedures
- 71% eventually develop lower extremity weakness and 30% develop upper extremity weakness
- CK is normal or mildly elevated
- Muscle biopsy shows rimmed vacuoles
- (GCN) trinucleotide repeat in the polyadenylation-binding protein nuclear gene 1 (PABPN1)

Oculopharyngeal muscular dystrophy (OPMD) is important to recognize because this can be mistaken for myasthenia gravis, as patients will come in with ptosis, extraocular muscle weakness, dysphagia, and often some facial weakness late in the course of the disease (Figure 41).

Therefore, OPMD has features of both MP5 (eyeball pattern) and MP7 (bulbar pattern). One of the clues favoring OPMD rather than MG is that because eye muscle weakness develops slowly throughout the entire lifetime in OPMD, the patient's eyes may not move at all, but they have no double vision. On the other hand, in myasthenia gravis, there may be no obvious deficit in extraocular motility on examination, but the patients complain of diplopia. Muscle biopsies may show the presence of rimmed vacuoles and subtle dystrophic features without inflammation. The definitive test demonstrates the genetic abnormality of a trinucleotide repeat expansion in the PABPN1 gene.

Limb-girdle muscular dystrophy (LGMD) (Figure 42)

Historically, limb-girdle muscular dystrophies were always lumped into one group until genetic mutations were found and there were apparent multiple limb-girdle muscular dystrophies. Most present with an MP1 pattern that is slowly progressive, however, distal presentations can occur (MP2) in some of the LGMDs.

Figure 42

Limb-Girdle Muscular Dystrophies

- Slowly progressive muscle weakness (MP1)
- Age of onset can range from childhood to 5-6th decades
- More than 20 known genetic mutations
- Other associated features can include
 - Skeletal involvement
 - CNS abnormalities
 - Some have distal weakness as well (MP2)
 - Cardio-respiratory involvement
- CK normal to 10,000s
- EMG with chronic myopathic and irritable changes
- Diagnosis by muscle biopsy and specific genetic testing

The most common forms of limb-girdle muscular dystrophy are shown in Figure 43.

Figure 43

Most Common Forms of Limb-Girdle MD

TYPE	GENE	PREVALENCE
LGMD 2A	Calpain 3	30%
LGMD 2B	Dysferlin	19%
LGMD 2I	Fukutin RP	18%
LGMD 1B	Lamin A/C, Emerin	12%
LGMD 2D	alpha-sarcoglycan	9%

Figure 44 provides more distinguishing features between a number of the LGMDs.

Figure 44

Distinguishing Features for Diagnosis of Limb-Girdle Muscular Dystrophies

Disease	Protein	Linkage (in years)	Age at Onset	Clinical Pearl	Early Distal Involvement	Cardiac Involvement	CK
LGMD1A	Myotilin	5q22.3-q13	20-40	Dysarthria	No	No	NL-10X
LGMD1B	Lamin A/C	1q13-q21	<10	Joint contractures	Sometimes	Yes	NL-20X
LGMD1C	Caveolin-3	3p25	5-25	Mounding / Rippling	Reported	No	2-25X
LGMD1D	Unknown	4p23	15-50	Cardiomyopathy	No	Yes	NL-4X
LGMD1E	Unknown	7q	30-50	-	No	No	NL-10X
LGMD1F	Unknown	7q32.1-q32.2	<15 & >20	Anticipation	No	No	NL-15X
LGMD1G	Unknown	4p21	30-47	Finger flexion limitation	Yes	No	NL-20X
LGMD2A	Calpain-3	15q15.1	5-40	Adductor weakness	No	No	NL-50X
LGMD2B	Dysferlin	2p13	10-30	Distal leg involvement	Yes	No	2-150X
LGMD2C	α, β, γ, δ-Sarcoglycan	13q32.1-q34.1	5-20	"Duchenne-like" No	Yes	Yes	5-120X
LGMD2D	Telethonin	17q11-q12	2-15	Brazilian	Yes	Yes	2-30X
LGMD2H	TRIM32	9q31-q34	5-30	Hutterite	No	No	NL-20X
LGMD2I	Fukutin related protein	15q13.3	1-40	Respiratory dysfunction	No	Yes	5-40X
LGMD2J	Titin	2q31	5-20	Finnish	No	No	NL-4X
LGMD2K	α-mannosidase-1	9q24.1	<5	Mental retardation	No	No	20-40X
LGMD2L	Anoctamin 5	11p13-p12	10-50	Thigh involvement	No	No	NL-30X
LGMD2M	Fukutin	9q31	<5	Steroid responsive	No	No	5-30X
LGMD2N	O-mannose (β1,2, N-acetylglucosaminyl) transferase	1p32	12	MIB traits	No	No	20-50X
LGMD2O	O-mannosyltransferase-2	14q24	<2	MIB traits	No	No	20-30X

References:
Barohn RJ. In: *Cecil Textbook of Medicine*, 23rd ed. Philadelphia, PA: Saunders Elsevier, 2008:2816-2834.
Amato AA, Russell JA. *Neuromuscular Disorders*. New York, NY: McGraw-Hill Companies, Inc.; 2016.

Congenital Muscular Dystrophy

Congenital muscular dystrophies occur within the first year of life and present with prominent hypotonia. Ultimately an MPI pattern is recognizable. Frequently other organ systems are involved including eyes, lungs, brain, and heart (Figure 45).

Figure 45

Congenital Muscular Dystrophy

- MP1
- Onset of weakness within the first year
- Prominent hypotonia
- Delayed development of motor skills
- Progressive weakness
- Often involves other organs including eyes, brains, lungs and heart




Figure 46 shows a muscle biopsy with dystrophic changes, muscle size variability, and significant replacement of muscle fibers by connective tissue. However, a muscle biopsy is now rarely indicated, and patients are diagnosed with specific genetic testing.

Figure 46

Congenital Muscular Dystrophies: Diagnosis

- Elevated CPK
- EMG with myopathic features
- Brain MRI to look for central nervous system involvement
- Muscle biopsy
- Specific genetic testing

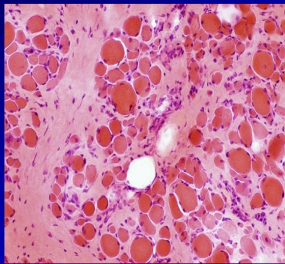


Figure 47 shows a number of different congenital muscular dystrophies with specific genetic abnormalities. While often included on this list, Bethlem myopathy is due to a genetic mutation in collagen genes rather than muscle-specific genes.

Figure 47

Congenital Muscular Dystrophies				
	Respiratory	Cardiac	Brain MRI	Course
Merosin	Common	35%	T2 Abnormal	Slowly progressive
Fukuyama	Not seen	Decreased EF	migrational defects	Progressive
Musce-Eye-Brain	Not seen	Not seen	Structural abnormal	Progressive
Walker-Warburg	Not seen	Not common	Structural abnormal	Poor survival
Bethlem	Not seen	some conduction defects	Normal	Progression at 2nd decade
Ullrich	Early resp involvement	Not seen	Normal	Progressive

Congenital Myopathies

There are several congenital myopathies that usually present in childhood with an MP1 pattern. By definition, these tend to be nonprogressive in contradistinction to congenital muscular dystrophies. A number of different molecular genetic defects have now been identified for nemaline rod, congenital myopathy with central nuclei, and congenital myopathy with cores.

Figure 48

Congenital Myopathies

- MP1, occasionally MP2, MP3, MP5
- Early onset hypotonia
- Muscle weakness
- Not progressive by definition
- NI to slightly elevated CPK
- Muscle biopsy with specific pathologic findings

Figure 49

**Congenital Myopathies:
Nemaline Rod**

- 90% are congenital but childhood and adult onset do occur
 - Can be autosomal dominant or recessive
 - 7 identified genes to date
- Weakness and hypotonia
- Cognitive involvement in younger children
- Progressive disorder in kids with death from respiratory involvement
- Adult can present with paraspinous and neck extensor involvement

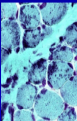


Figure 50

Congenital Myopathies

- Congenital Myopathy with Fiber Size Disproportion
- Autosomal dominant or recessive
- Onset is congenital or childhood
- Diffuse weakness and hypotonia
- 50% are static, 35% improve somewhat and 15% worsen

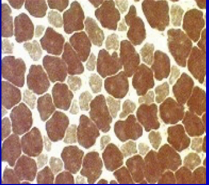


Figure 51

Congenital Myopathies

- Congenital myopathy with central nuclei
 - X-linked (Myotubularin 1) or autosomal dominant or recessive
 - Onset in infancy
 - 50% of mothers will have polyhydramnios
 - Facial and head malformations
 - Hypotonia, muscle and respiratory weakness and ocular (ptosis, EOM)
 - Many die within the first 6 months
 - Longer survivors require respiratory support

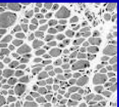
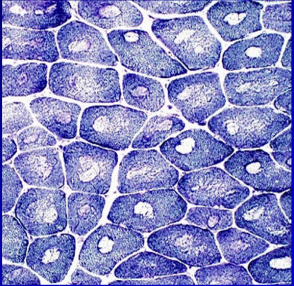


Figure 52

**Congenital Myopathies:
CORES**

- Ryanodine receptor syndromes
- Central core myopathy
- Congenital or childhood onset
- Autosomal dominant or recessive
- Hypotonia
- Congenital hip dislocation and reduced fetal movements
- Slowly or not progressive
- Malignant hyperthermia
- Usually Limb Girdle
- MP1



Metabolic Myopathies

The last group of disorders we will consider in this lecture are the metabolic myopathies. We have already discussed Pompe disease which is included in this group and is considered a static metabolic myopathy. Another group of metabolic myopathies with static presentation

are the mitochondrial myopathies which can be associated with not only an MP1 presentation but also the eyeball presentation and ptosis with ophthalmoplegia (MP5). Other metabolic myopathies are episodic and present with rhabdomyolysis and myoglobinuria, MP8 pattern (Figure 53). These can also be considered dynamic disorders. Metabolic myopathies that have exercise intolerance and rhabdomyoma provoked by intense exercise under 10 minutes are usually glycogen disorders. On the other hand, dynamic metabolic myopathies in which symptoms are provoked by low-intensity exercise lasting longer than 10 minutes are usually lipid disorders.

Figure 53

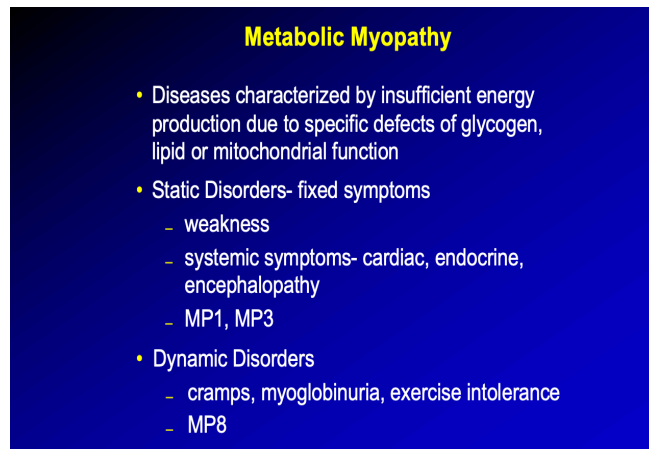


Figure 54

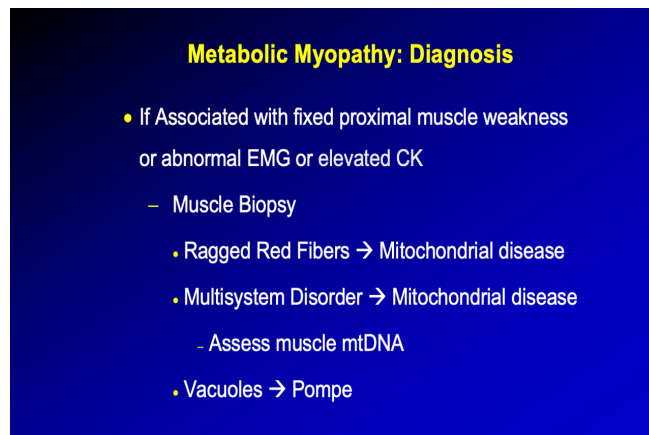


Figure 55

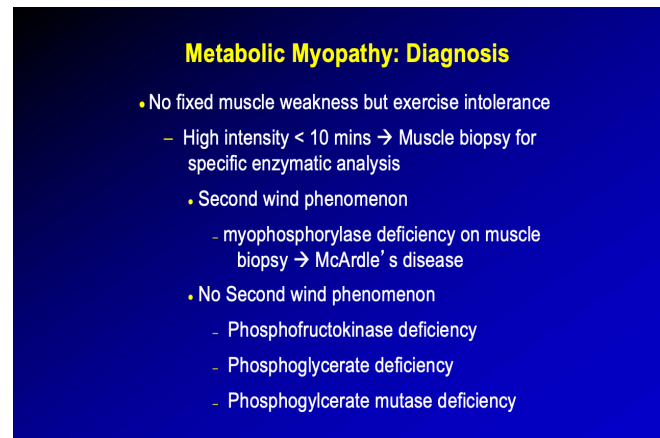
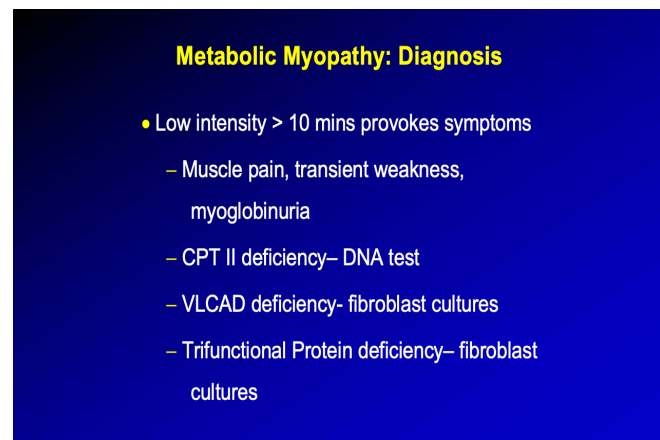


Figure 56



We would like to thank Amanda Sebok for her assistance in preparing the PowerPoint figures and Lauren Peck for her editorial assistance in preparing the manuscript.

References

- Amato AA, Gronseth GS, Jackson CE, Wolfe GI, Katz JS, Bryan WW, Barohn RJ. Inclusion body myositis: clinical and pathological boundaries. *Ann Neurol*. 1996 Oct;40(4):581-6.
- Amato AA, Barohn RJ. Evaluation and treatment of inflammatory myopathies. *J Neurol Neurosurg Psych*. 2009; 80(10):1060-1068.
- Amato AA, Barohn RJ. Inclusion body myositis: old and new concepts. *J Neurol Neurosurg Psych*. 2009; 80(11):1186-1193.
- Amato AA, Tawil R, Kissel J, Barohn RJ, McDermott MP, Pandya S, King W, Smirnow A, Annis C, Roe K, Tawil R, Janciuras J, Dilek N, Martens WB, Eastwood E, Cochrane T, Donlan M, Chused S, Dimachkie MM, Aires DJ, Latinis KM, Herbelin L, Michaels H, Cupler E, Deodhar A, Simpson E, Burusnukul P, Edgar E, Serdar A, Brennan T, Gance K, Freimer ML, Hackshaw KV, Lawson V, King WM, Bartlett A, Wolfe G, Nations S, McLin R, Gorham N, Briemberg H, Chapman KM, Dutz JP, Wilson J, Varelas F, Wagner K, Stine LC, Anhalt GJ, Meyerle JH, Swain JO, Brock-Simmons R, Weiss M, Distad BJ, Lin J, Haug JA, Downing S. A randomized, pilot trial of etanercept in dermatomyositis. *Ann Neurol*. 2011; 70(3):427-436.
- Amato AA, Doughty CT, Paganoni S, Guidon AC. Amato and Russell's Neuromuscular Disorders. 3rd Edition. New York. McGraw-Hill Education; 2025.
- Barohn RJ, Dimachkie MM, Jackson CE. A pattern recognition approach to patients with a suspected myopathy. *Neurol Clin*. 2014; 32(3):569-593.
- Baraliakos X, Ranza R, Östör A, Ciccio F, Coates LC, Rednic S, Walsh JA, Douglas K, Gao T, Kato K, Song IH, Ganz F, Deodhar A. Efficacy and safety of upadacitinib in patients with active psoriatic arthritis and axial involvement: results from two phase 3 studies. *Arthritis Res Ther*. 2023; 25(1):56.
- Bottai M, Lundberg IE, Tjärnlund A, Werth VP, Pilkington C, Visser M, Alfredsson L, Amato AA, Barohn RJ, Liang MH, Singh JA, Aggarwal R, Arnardottir S, Chinoy H, Cooper RG, Dankó K, Dimachkie MM, Feldman BM, Torre IG, Gordon P, Hayashi T, Katz JD, Kohsaka H, Lachenbruch PA, Lang BA, Li Y, Oddis CV, Olesinska M, Reed AM, Rutkowska-Sak L, Sanner H, Selva-O'Callaghan A, Song YW, Vencovsky J, Ytterberg SR, Miller FW, Rider LG. International Myositis Classification Criteria Project consortium, The Euromyositis register and The Juvenile Dermatomyositis Cohort Biomarker Study and Repository (JDRG) (UK and Ireland). 2017 EULAR/ACR Classification Criteria for Adult and Juvenile Idiopathic Inflammatory Myopathies and their major subgroups. *Ann Rheum Dis*. 2017;76(12):1955-1964.
- Chahin N, Engel AG. Correlation of muscle biopsy, clinical course, and outcome in PM and sporadic IBM. *Neurology*. 2008; 70(6):418-424.
- Christopher-Stine L, Casciola-Rosen LA, Hong G, Chung T, Corse AM, Mammen AL. A novel autoantibody recognizing 200-kd and 100-kd proteins is associated with an immune-mediated necrotizing myopathy. *Arthritis Rheum*. 2010; 62(9):2757-2766.
- Dasouki M, Jawdat O, Almadhoun O, Pasnoor M, McVey AL, Abuzinadah A, Herbelin L, Barohn RJ, Dimachkie MM. Pompe disease: literature review and case series. *Neurol Clin*. 2014; 32(3):751-776.
- Dimachkie MM, Barohn RJ, Amato AA. Idiopathic inflammatory myopathies. *Neurol Clin*. 2014; 32(3):595-628.
- Dimachkie MM, Barohn RJ. Inclusion body myositis. *Neurol Clin*. 2014; 32(3):629-646.
- Dimachkie MM, Barohn RJ. Distal myopathies. *Neurol Clin*. 2014; 32(3):817-842.
- Dimachkie MM, Barohn RJ, Byrne B, Goker-Alpan O, Kishnani PS, Ladha S, Laforêt P, Mengel KE, Peña LDM, Sacconi S, Straub V, Trivedi J, Van Damme P, van der Ploeg AT, Vissing J, Young P, Haack KA, Foster M, Gilbert JM, Miossec P, Vitse O, Zhou T, Schoser B. Long-term Safety and Efficacy of Avalglucosidase Alfa in Patients With Late-Onset Pompe Disease. *Neurology*. 2022; 99(5):536-548.
- Flanigan KM. Duchenne and Becker muscular dystrophies. *Neurol Clin*. 2014; 32(3): 671-688.
- Gilbreath HR, Castro D, Iannaccone ST. Congenital myopathies and muscular dystrophies. *Neurol Clin*. 2014; 32(3):689-703.
- Guglieri M, Bushby K, McDermott MP, Hart KA, Tawil R, Martens WB, Herr BE,
- McColl E, Speed C, Wilkinson J, Kirschner J, King WM, Eagle M, Brown MW, Willis T, Griggs RC, Straub V, van Ruiten H, Childs AM, Cialfoni E, Shieh PB, Spinty S, Maggi L, Baranello G, Butterfield RJ, Horrocks IA, Roper H, Alhaswani Z, Flanigan KM, Kuntz NL, Manzur A, Darras BT, Kang PB, Morrison L, Krzesniak-Swinarska M, Mah JK, Mongini TE, Ricci F, von der Hagen M, Finkel RS, O'Reardon K, Wicklund M, Kumar A, McDonald CM, Han JJ, Joyce N, Henricson EK, Schara-Schmidt U, Gangfuss A, Wilichowski E, Barohn RJ, Statland JM, Campbell C, Vita G, Vita GL, Howard JF Jr, Hughes I, McMillan HJ, Pegoraro E, Bello L, Burnette WB, Thangarajh M, Chang T. Effect of Different Corticosteroid Dosing Regimens on Clinical Outcomes in Boys With Duchenne Muscular Dystrophy: A Randomized Clinical Trial. *JAMA*. 2022; 327(15):1456-1468.
- Hill CL, Zhang Y, Sigurgeirsson B, Pukkala E, Mellemkjaer L, Airio A, Evans SR, Felson DT. Frequency of specific cancer types in dermatomyositis and polymyositis: a

- population-based study. *Lancet*. 2001; 357(9250):96-100.
- Jabari D, Vedanarayanan W, Barohn RJ, Dimachkie MM. Update on Inclusion Body Myositis. *Curr Rheumatol Rep*. 2018; 28;20(8):52.
- Jackson CE, Barohn RJ. Improvement of the exercise test after therapy in thyrotoxic periodic paralysis. *Muscle Nerve*. 1992; 15(10):1069-1071.
- Kishnani PS, Corzo D, Nicolino M, Byrne B, Mandel H, Hwu WL, Leslie N, Levine J, Spencer C, McDonald M, Li J, Dumontier J, Halberthal M, Chien YH, Hopkin R, Vijayaraghavan S, Gruskin D, Bartholomew D, van der Ploeg A, Clancy JP, Parini R, Morin G, Beck M, De la Gastine GS, Jokic M, Thurberg B, Richards S, Bali D, Davison M, Worden MA, Chen YT, Wraith JE. Recombinant human acid [α]-glucosidase: major clinical benefits in infantile-onset Pompe disease. *Neurology*. 2007 Jan 9;68(2):99-109.
- Logigian EL, Martens WB, Moxley RT, McDermott MP, Dilek N, Wiegner AW, Pearson AT, Barbieri CA, Annis CL, Thornton CA, Moxley RT. Mexiletine is an effective antimyotonia treatment in myotonic dystrophy type 1. *Neurology*. 2010; 74(18):1441-1448.
- Machado PM, McDermott MP, Blaettler T, Sundgreen C, Amato AA, Cifaloni E, Freimer M, Gibson SB, Jones SM, Levine TD, Lloyd TE, Mozaffar T, Shaibani AI, Wicklund M, Rosholm A, Carstensen TD, Bonefeld K, Jørgensen AN, Phonekeo K, Heim AJ, Herbelin L, Barohn RJ, Hanna MG, Dimachkie MM; Arimocloleol in IBM Investigator Team of the Neuromuscular Study Group. Safety and efficacy of arimocloleol for inclusion body myositis: a multicentre, randomised, double-blind, placebo-controlled trial. *Lancet Neurol*. 2023 Oct;22(10):900-911.
- Mammen AL, Chung T, Christopher-Stine L, Rosen P, Rosen A, Doering KR, Casciola-Rosen LA. Autoantibodies against 3-hydroxy-3-methylglutaryl-coenzyme A reductase in patients with statin-associated autoimmune myopathy. *Arthritis Rheum*. 2011; 63(3):713-721.
- Mammen AL, Pak K, Williams EK, Brisson D, Coresh J, Selvin E, Gaudet D. Rarity of anti-3-hydroxy-3-methylglutaryl-coenzyme A reductase antibodies in statin users, including those with self-limited musculoskeletal side effects. *Arthritis Care Res (Hoboken)*. 2012; 64(2):269-272.
- McManis PG, Lambert EH, Daube JR. The exercise test in periodic paralysis. *Muscle Nerve*. 1986; 9(8):704-710.
- Mendell JR, Muntoni F, McDonald CM, Mercuri EM, Cifaloni E, Komaki H, Leon-Astudillo C, Nascimento A, Proud C, Schara-Schmidt U, Veerapandian A, Zaidman CM, Guridi M, Murphy AP, Reid C, Wandel C, Asher DR, Darton E, Mason S, Potter RA, Singh T, Zhang W, Fontoura P, Elkins JS, Rodino-Klapac LR. AAV gene therapy for Duchenne muscular dystrophy: the EMBARK phase 3 randomized trial. *Nat Med*. 2024; 31(1):332-341.
- Morgan C, Nozaki K, Park DJ, Pestronk A, Rosenbloom B, Skrinar A, van Capelle CI, van der Beek NA, Wasserstein M, Zivkovic SA. A randomized study of alglucosidase alfa in late-onset Pompe's disease. *N Engl J Med*. 2010; 362(15):1396-1406.
- Naddaf E, Barohn RJ, Dimachkie MM. Inclusion Body Myositis: Update on Pathogenesis and Treatment. *Neurotherapeutics*. 2018; 15(4):995-1005.
- Needham M, Fabian V, Knezevic W, Panegyres P, Zilko P, Mastaglia FL. Progressive myopathy with up-regulation of MHC-I associated with statin therapy. *Neuromuscul Disord*. 2007; 17(2):194-200.
- Pasnoor M, Khan S, Jackson CE, Kissel J, Wolfe GI, Nations SP, Trivedi JR, Singer MA, Elliott JL, Vernino S, Mummaneni RB, Herbelin L, Saperstein DS, Gronseth G, Mcvey A, Dimachkie MM, Barohn RJ. Database Evaluation for Muscle and Nerve Diseases - DEMAND: An academic neuromuscular coding system. *RRNMF Neuromuscular Journal*. 2023;4(4):45-67.
- Pasnoor M, Barohn RJ, Dimachkie MM. Toxic myopathies. *Neurol Clin*. 2014; 32(3):647-670.
- Pluk H, van Hoeve BJ, van Dooren SH, Stammen-Vogelzangs J, van der Heijden A, Schelhaas HJ, Verbeek MM, Badrising UA, Arnardottir S, Gheorghe K, Lundberg IE, Boelens WC, van Engelen BG, Puijn GJ. Autoantibodies to cytosolic 5'-nucleotidase 1A in inclusion body myositis. *Ann Neurol*. 2013 Mar;73(3):397-407.
- Rockette HE. Rituximab in Myositis (RIM) Study Group. Rituximab in the treatment of refractory adult and juvenile dermatomyositis and adult polymyositis: A randomized, placebo phase trial. *Arthritis and Rheumatism*. 2013; 65(2):314-324.
- Salajegheh M, Lam T, Greenberg SA. Autoantibodies against a 43 KDa Muscle Protein in Inclusion Body Myositis. *PLOS ONE*. 2011; 6(5).
- Sharp LJ, Haller RG. Metabolic and mitochondrial myopathies. *Neurol Clin*. 2014; 32(3):777-799.
- Statland JM, Bundy BN, Wang Y, Rayan DR, Trivedi JR, Sansone VA, Salajegheh MK, Venance SL, Cifaloni E, Matthews E, Meola G, Herbelin L, Griggs RC, Barohn RJ, Hanna MG. Mexiletine for symptoms and signs of myotonia in nondystrophic myotonia: a randomized controlled trial. *JAMA*. 2012; 308(13):1357-1365.
- Statland JM, Tawil R. Facioscapulohumeral Muscular Dystrophy. *Continuum (Minneapolis)*. 2014; 22(6):1916-1931.
- Statland J, Phillips L, Trivedi JR. Muscle channelopathies. *Neurol Clin*. 2014; 32(3):801-815.
- Thornton CA. Myotonic dystrophy. *Neurol Clin*. 2014; 32(3):705-719.

- Trivedi JR, Bundy B, Statland J, Salajegheh M, Rayan DR, Venance SL, Wang Y, Fialho D, Matthews E, Cleland J, Gorham N, Herbelin L, Cannon S, Amato AA, Griggs RC, Hanna MG, Barohn RJ. Non-dystrophic myotonia: prospective study of objective and patient reported outcomes. *Brain*. 2013; 136(Pt 7):2189-2200.
- van der Ploeg AT, Clemens PR, Corzo D, Escolar DM, Florence J, Groeneveld GJ, Herson S, Kishnani PS, Laforet P, Lake SL, Lange DJ, Leshner RT, Mayhew JE, Valiyil R, Casciola-Rosen L, Hong G, Mammen A, Christopher-Stine L. Rituximab therapy for myopathy associated with anti-signal recognition particle antibodies: a case series. *Arthritis Care Res (Hoboken)*. 2010; 62(9):1328-1334.
- van der Meulen MF, Bronner IM, Hoogendijk JE, Burger H, van Venrooij WJ, Voskuyl AE, Dinant HJ, Linssen WH, Wokke JH, de Visser M. Polymyositis: an overdiagnosed entity. *Neurology*. 2003; 61(3):316-321.
- Vivekanandam V, Skorupinska I, Jayaseelan DL, Matthews E, Barohn RJ, McDermott MP, Hanna MG. Mexiletine versus lamotrigine in non-dystrophic myotonias: a randomized, double-blind, head-to-head, crossover, non-inferiority, phase 3 trial. *Lancet Neurol*. 2024; 23(10):1004-1012.
- Walton JN, Nattrass FJ. On the classification, natural history and treatment of the myopathies. *Brain*. 1954; 77(2):169-231.
- Walton JN, Nattrass FJ. On the classification, natural history and treatment of the myopathies. *Brain*. 1954; 77(2):169-231.
- Walton JN, Adams RD. Polymyositis. Williams & Wilkins. 1958;100-105.
- Wicklund MP, Kissel JT. The limb-girdle muscular dystrophies. *Neurol Clin*. 2014; 32(3):729-749.
- Wilson FC, Ytterberg SR, St Sauver JL, Reed AM. Epidemiology of sporadic inclusion body myositis and polymyositis in Olmsted County, Minnesota. *J Rheumatol*. 2008; 35(3):445-447.