



Dual-specificity tyrosine-regulated kinases (DYRKs) and cancer

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ABSTRACT

Dual-specificity tyrosine phosphorylation-regulated kinase (DYRK) belongs to the CMGC group of kinases that are named after cyclin-dependent kinases (CDKs), mitogen-activated protein kinases (MAPKs), glycogen synthase kinases (GSKs), and CDC-like kinases (CLKs). DYRK-related kinases comprise a novel subfamily of protein kinases with unique structural, biochemical, and enzymatic features. In humans, DYRKs phosphorylate a set of proteins and play a critical role in apoptosis, DNA damage repair, cell proliferation, survival, and development. Dysregulation of DYRK protein kinases have been associated with cancer biology. In recent years, several studies have reported some kinase inhibitors affecting cancer development and progression, making them potential therapeutic drugs. However, challenges remain in understanding the molecular mechanisms and roles of each member of the DYRK family in cancer initiation and progression. In this review, we will highlight the importance of DYRK kinases in cancer biology.

ARTICLE HISTORY

Received: Nov 8, 2024
Revised: Nov 27, 2024
Accepted: Nov 28, 2024

KEYWORDS

DYRK, kinase, phosphorylation, signaling pathway, cancer

EDITED BY

Qianben Wang

REVIEWED BY

Zhiguo Li, Shang Su

Background

Cancer is an abnormal growth of cells. Cancer cells have mutations that lead to dysregulated expression of oncogenes and tumor suppressor genes, the results of these changes altering the expression of hundreds of genes [1-3]. The proto-oncogene c-Src is the first cancer gene identified to encode a protein kinase [4]. In human cells, 538 protein kinases have been identified [5]. Based on the sequence homology of their catalytic domains, these protein kinases are classified into several groups [6]. Dual-specificity tyrosine phosphorylation-regulated kinases (DYRKs) are a subfamily of the CMGC kinase group that encompasses cyclin-dependent kinases (CDKs), mitogen-activated protein kinases (MAPKs), glycogen synthase kinases (GSKs), and CDK-like kinases (CLKs), as well as tyrosine kinase gene v-ros cross hybridizing kinases (RCKs) [6]. The extended DYRK family members are classified into three subfamilies: Dual specificity (DYRK) kinases, pre-messenger RNA processing protein 4 kinases (PRP4s), and

homeodomain-interacting protein kinases (HIPKs) [6]. The DYRK subfamily is clustered into two classes (Class I and Class II) with five members in humans. Class I DYRKs include DYRK1A and DYRK1B (also known as Mirk from mini brain-related kinase), and Class II has DYRK2, DYRK3, and DYRK4.

The DYRK members are autophosphorylated on a tyrosine residue to reach full kinase activities [7, 8] and play key roles in signaling pathways that control cell survival, differentiation, cell death, embryonic neurogenesis, development processes [9, 10], and cancer biology [11]. Regarding DYRK involvement in the regulation of tumorigenic processes, several studies have reported alterations in the expression of DYRK genes in tumor samples and have revealed DYRK-dependent mechanisms that contribute to tumor initiation and progression. However, challenges remain, including DYRK kinase inhibitors, clinical implications, and therapeutic opportunities. This review will highlight the importance of DYRK kinases in their involvement in signaling pathways and cancer biology.

DYRK kinase subfamily

DYRK-related kinases compose a distinct family of protein kinases defined by the structural similarity of their kinase domains and their capability to autophosphorylate on tyrosine residues. Autophosphorylation of DYRKs on tyrosine is a one-off event that occurs during translation and induces kinase activation [12, 13]. The members of the DYRK family have unrelated sequences outside the catalytic kinase domain (DH-box, Dyrk homology box), and differ in their tissue distribution, substrate specificity, and subcellular localization [14]. Class I DYRKs contain a functional, bipartite nuclear localization signal (NLS) at the N-terminus to the DH-box, and a proline-, glutamic acid-, serine-, and threonine-rich (PEST) at the C-terminus which is typically found in proteins that are targeted for degradation. Class I also contains a histidine-rich region in the C terminus [11, 15]. Class II includes a conserved kinase domain and adjacent N-terminal autophosphorylation accessory region (NAPA) domain to the DH box but not a PEST domain at the C-terminus (Figure 1) that is essential for autophosphorylation of the activation loop tyrosine [16].

The kinase structural domain of DYRK1A contains an N-terminal lobe (N-lobe) with five antiparallel β -strands, a conserved regulatory α C helix, and a larger C-terminal lobe (C-lobe) [17]. DYRK1A and DYRK1B are closely related and highly expressed in

human tissues [11], and both are characterized as negative regulators of the cell cycle and are involved in cell survival, growth arrest, and differentiation [18, 19]. The mammalian mini-brain (MNB) ortholog DYRK1A plays a key role in the MNB-like phenotype and aberrant retinal development [20, 21], which indicates a conserved mechanism for the normal development of the central nervous system in mammals [16]. Moreover, DYRK1A protein kinase has a nonredundant vital role in the maintenance of adult brain neuronal activities by regulating transcription factor, nuclear factor of activated T cells (NFAT) [9], and cyclin AMP response element-binding protein (CREB) [22]. DYRK1A and DYRK3-specific runs of histidine residues participate in phase-separated subcellular compartments [23]. DYRK1B has been illustrated as a negative regulator of the cell cycle and encodes a dual-specificity serine/threonine (S/T) protein kinase with roles in cell survival and differentiation [24, 25]. In addition, the kinase activity of DYRK1B attenuates DNA double-strand breaks (DSBs)-induced gene silencing and leads to compromised DNA damage repair that coordinates DSB repair on transcribed chromatin [26]. DYRK2 phosphorylates NFATc and regulates calcium signaling, resulting in NFATc inactivation through cytoplasmic sequestration [9, 27]. Recent studies have shown that DYRK2 is a novel mammalian ciliogenesis-related protein kinase that plays an important role in regulating Hedgehog (Hh) signaling in the control of ciliogenesis [28]. DYRK2

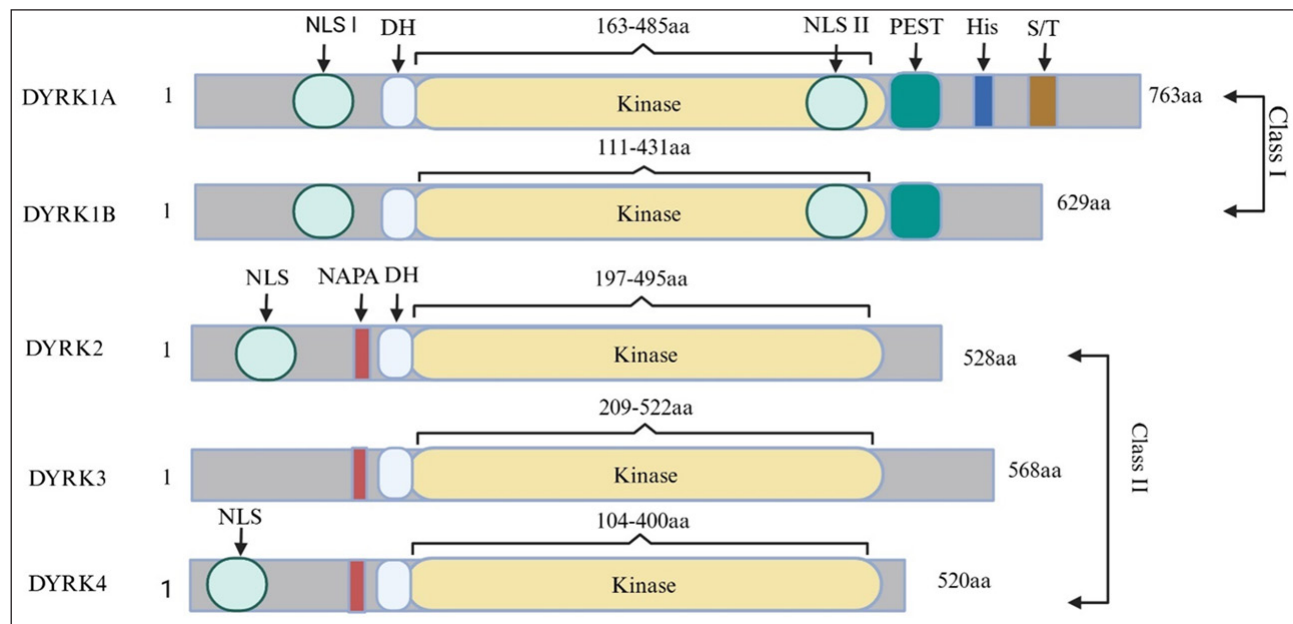


Figure 1: Schematic representation of the mammalian family of DYRKs, indicating their phylogenetic relationships, degree of homology, and protein domains. NLS: nuclear localization; DH: DYRK homology; PEST: proline-, glutamic acid-, serine-, and threonine-rich; His: histidine; S/T: serine/threonine; aa: amino acid. Image created in BioRender. Moududee, S. (2024) <https://BioRender.com/k39x947>.

and DYRK3 are closely related and encoded by paralogous genes that originate from gene duplication [29]. DYRK3 is localized to stress granules (SG) and regulates a cyclic partitioning mechanism between SG and cytosol through its kinase activity [30]. DYRK1A, DYRK2, and DYRK4 differ in their substrate specificities whereas DYRK2 and DYRK4 are less biased than DYRK1A. DYRK4 is widely expressed in all human tissues, unlike its rat and mouse orthologues that are predominantly expressed in the testis [31]. In neurons, DYRK2, DYRK3, and DYRK4 are mostly localized in the cytosolic portion of cell bodies, with DYRK2 seen in the dendrites and part of the axon and DYRK3 and DYRK4 mainly confined to the dendrites; in contrast, DYRK1A is mainly localized to the nucleus [32]. Taken altogether, the structural differences and subcellular localizations of DYRKs may determine their differential functions in the cells.

DYRKs in Cancer

Protein phosphorylation is one of the most significant mechanisms for signal transduction between cells and within cells. DYRK phosphorylation can influence a broad range of cellular processes including

transcription, apoptosis, cell cycle progression, cell motility, metabolism, cell survival, and differentiation, which play important roles in human diseases including various cancers. DYRK family members have been linked to cancer biology as their expression is different between normal and tumor tissues and many DYRK substrate proteins are key players in cancer biology [11]. In this section, we will discuss the involvement of each member of the DYRK family in cancer.

DYRK1A

The gene encoding DYRK1A is highly conserved and located on chromosome 21 in the Down syndrome critical region (DSCR) [33]. Previous studies have illustrated that individuals with Down syndrome show a markedly reduced incidence of solid tumors [34] and a considerably lower incidence of cancer-associated mortality [35]. DYRK1A is a pleiotropic kinase that phosphorylates proteins that play crucial roles in angiogenesis, DNA damage repair, cell cycle regulation, cancer stem cell properties, transcription, and cell signaling regulation [36] (Figure 2). DSCR1 is located in the Down syndrome critical region

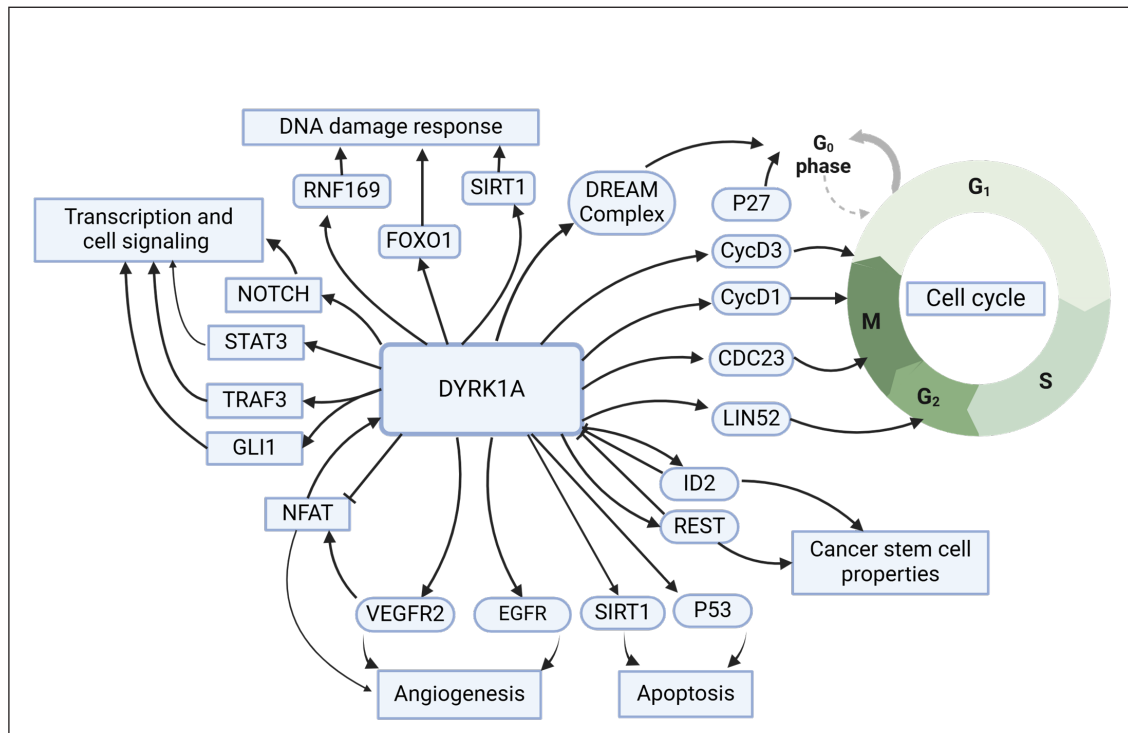


Figure 2: An overview of interactions between DYRK1A and proteins involved in a variety of cellular processes. RNF169: Ring finger protein 169; FOXO1: Forkhead box protein 1; SIRT1: silent information regulator sirtuin 1; NOTCH: neurogenic locus notch homolog protein; STAT3: signal transducer and activator of transcription 3; TRAF3: TNF receptor-associated factor; GLI1: glioma-associated oncogene 1; NFAT: nuclear factor of activated T-cells; VEGFR2: Vascular endothelial growth factor receptor 2; EGFR: epidermal growth factor receptor; p53: tumor protein 53; REST: RE1-silencing factor; ID2: inhibitor of DNA Binding 2; LIN52: protein lin-52 homolog; CDC23: Cell division cycle 23; CycD1: cyclin D1; CycD3: cyclin D3; DREAM: dimerization partner (DP), retinoblastoma (RB)-like, E2F and MuvB complex. Image created in BioRender. Moududee, S. (2024) <https://BioRender.com/k39x947>.

and coordinates with DYRK1A to reduce angiogenesis through vascular endothelial growth factor 2 (VEGF2)-dependent signaling and the downstream nuclear factor of activated T cells (NFAT)-dependent transcriptional response in endothelial cells [9, 37]. DYRK1A contributes to acute megakaryoblast leukemia (AMKL) development through NFAT signaling [38]. The signal transducer and activator of transcription 3 (STAT3) is a DYRK1A substrate that regulates tumor progression [39, 40] and astroglialogenesis [41]. STAT3 is activated in solid tumors such as lung, prostate, breast, head, neck, gastric, colorectal, hepatocellular, and brain cancers [42]. The inhibition of DYRK1A diminishes STAT3 activities and proliferation of non-small cell lung cancer (NSCLC) cells due to impaired EGFR signaling [43].

Glioma-associated oncogene 1 (GLI1, officially named GLI Family Zinc Finger 1) is an oncogenic transcription factor whose nuclear translocation and functional activity are regulated through phosphorylation by DYRK1A. GLI1 is the effector of Sonic Hedgehog (SHH) signaling that regulates tumor growth and cell survival as well as metastasis [44]. Moreover, the NOTCH pathway has appeared as an oncogenic pathway in solid tumors and blood cancers [45]. On the other hand, it has also been revealed

that the NOTCH pathway plays a role as a tumor suppressor in myeloid malignancies [46]. These findings suggest that the NOTCH pathway may have both positive and negative impacts on tumor growth and cell survival. The RE1-silencing factor (REST) is like NOTCH and has been identified as a cancer-related gene in medulloblastoma [47, 48], characterized as an oncogene in neuroblastoma (NB), a common extracranial solid tumor in children [49], and glioblastoma (GBM) that is in the adult brain [50, 51]. Therefore, DYRK1A has diverse, non-redundant roles in cancer biology, which may be a potential target in developing new strategies against cancers.

DYRK1B

DYRK1B is also named Minibrain-related kinase (Mirk). DYRK1B's functions have been well characterized. DYRK1B plays a vital role in muscle differentiation through regulatory effects on transcription, growth arrest, myogenesis, cell cycle progression, motility, and cell survival [25]. Thus, we will discuss the protumorigenic roles of DYRK1B in cancer, which mostly involve prosurvival signaling pathways (Figure 3). DYRK1B kinase is a potential pharmacological target in cancer since it is overexpressed in several

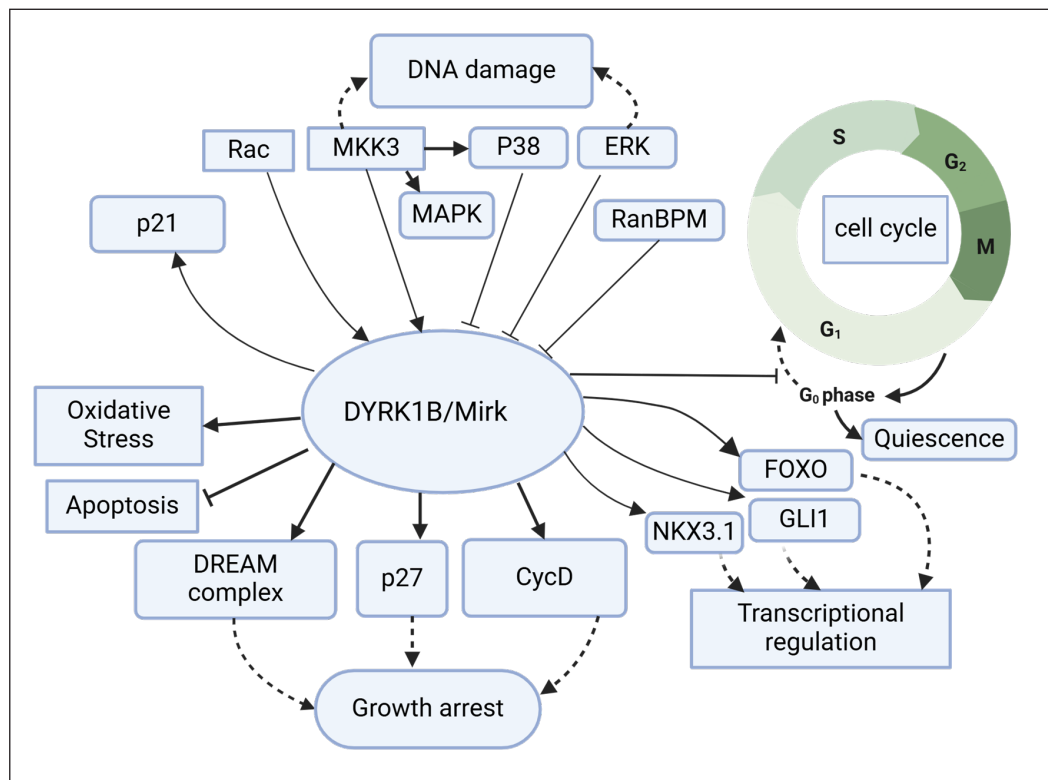


Figure 3: A graphical representation of the interactions between DYRK1B/Mirk and a variety of biological processes. MKK3: mitogen-activated protein kinase (MAPK) kinase 3; Rac: RAS-related C3 botulinum toxin substrate; ERK: extracellular signal-regulated kinase; RanBPM: Ran-binding protein M. CycD: Cyclin D; FOXO: Forkhead box protein; GLI1: glioma-associated oncogene 1. Image created in BioRender. Moududee, S. (2024) <https://BioRender.com/k39x947>.

types of solid tumors and cancer cell lines, including rhabdomyosarcoma [25], ovarian cancer [52], non-small cell lung carcinoma [53], breast cancer [54], prostate cancer, colon cancer [55, 56], and pancreatic ductal adenocarcinoma [57, 58], and its overexpression is correlated with patients' poor prognosis.

DYRK1B acts as a negative cell cycle regulator to maintain the survival of quiescent cancer cells [3, 59] and as a tumor cell survival factor conferring resistance to chemotherapies by counteracting the G0 / G1-S phase transition [60]. DYRK1B maintains the S phase checkpoint by stabilizing the cyclin-dependent kinase (CDK) inhibitor and induces the degradation of cyclin D [3], respectively. Additionally, the mammalian dimerization partner (DP), retinoblastoma (RB)-like, E2F and MuvB (DREAM) complex represses cell cycle-dependent genes during quiescence complex that represses cell cycle gene expression in G0-arrested cells [3, 19]. The regulation of the cell cycle by DYRK1B is mediated through cyclin D phosphorylation [61-63], suggesting that cancer cells require active Mirk/DYRK1B to maintain quiescent status that is less sensitive to chemotherapy and radiotherapy [64]. In this regard, pharmacological inhibition of DYRK1B may be a promising therapeutic approach to sensitize cancer cells to chemotherapy and radiotherapy.

Inhibition or depletion of DYRK1B has been shown to exaggerate DNA damage and apoptosis caused by chemotherapeutic drugs that target proliferating cells [65-68] in cancer cell lines of various tumor origins [69, 70]. Depletion of DYRK1B increases the nuclear translocation of Forkhead box protein O (FOXO) and enhances the apoptosis of ovarian cancer cells [52]. The functional interaction of DYRK1B and oncogenic transcription factor glioma-associated oncogene family zinc finger 1 (GLI1) significantly reduces *in vivo* tumor growth of GLI1-dependent pancreatic cancer cell lines [71]. In addition, DYRK1B has been reported to target the tumor suppressor NKX3.1 for proteasomal degradation in prostate cancer cell [72]. DYRK1B inhibition in prostate cancer can increase the NKX3.1 suppressor protein level besides its impact on cell cycle regulators [3]. However, it remains elusive to understand its mechanism fully. DYRK1B kinase is active in pancreatic and rhabdomyosarcoma cancer cells through oncogenic mitogen-activated protein kinase 3 (MKK3) signaling pathway [25, 57, 73]. In addition, MKK3 activates p38 in response to stress signals induced by DNA damage [73]. In conclusion, DYRK1B may be a potential pharmacological target in cancer therapy.

DYRK2

DYRK2 has been found in all eukaryotes, and surprisingly across all orthologues, the conserved function of DYRK2 is to regulate cell division and tissue development [74, 75]. Over the past two decades, many studies have found that the substrates of DYRK2 are involved in cell growth, cell survival, cell proliferation, development, gene transcription, and proteasomal regulation [24, 76, 77]. The role of DYRK2 in cancer is the focus of this review.

DYRK2 plays diverse roles as a tumor suppressor and an oncogene in different cancers such as breast, liver, colorectal, ovarian, gastric, lung, and prostate cancers, as well as in chronic myeloid leukemia [76, 78] (Figure 4). Yoshida et al have reviewed the tumor suppressor role of DYRK2 in different cancers with antitumorigenic functions involving apoptosis, cell cycle regulation, cancer stemness, epithelial-to-mesenchymal transition (EMT), and metastasis [79]. DYRK2 suppresses cancer cell proliferation and invasion by regulating cyclin-dependent kinase 14 (CDK14) expression [80]. Previous studies have reported that CDK14 is an oncogene [81-84], and upregulated expression of CDK14 promotes tumor cell proliferation, migration, and invasion in breast cancer [80, 81]. To elucidate the function of DYRK2 as an oncogene, some oncogenic substrates have been reported such as 26S proteasome [74, 85, 86], p53 [27, 87], Heat shock factor 1 (HSF1) [27], c-Myc [88], Seven in absentia homolog 2 (SIAH2) [89], and NOTCH1 [90]. DYRK2 phosphorylates and activates 26S proteasome and HSF1, thereby stimulating the proteotoxic stress pathway during tumorigenesis in breast cancer [78]. Cancer cells are exposed to proteotoxic stress by increasing protein folding capacity that is controlled by HSF1 or expanding the misfolded/aggregated protein degradation through 26S proteasome [91, 92]. DYRK2 phosphorylates the regulatory particle 3 (Rpt3) subunit on the ATPase ring of the 19S subunit of the proteasome on a conserved Thr25 site, leading to enhanced substrate translocation and degradation [74]. Expression of DYRK2 is inversely associated with Snail expression but positively associated with E-cadherin expression, hence DYRK2 regulates Snail and promotes epithelial-mesenchymal transition (EMT) to drive invasion in human breast cancer [93]. Ectopic expression of DYRK2 phosphorylates mTOR at Thr631, leading to ubiquitination and degradation [94]. The correlation between DYRK2 and mTOR regulates the sensitivity to Everolimus (an mTOR inhibitor) in hormone

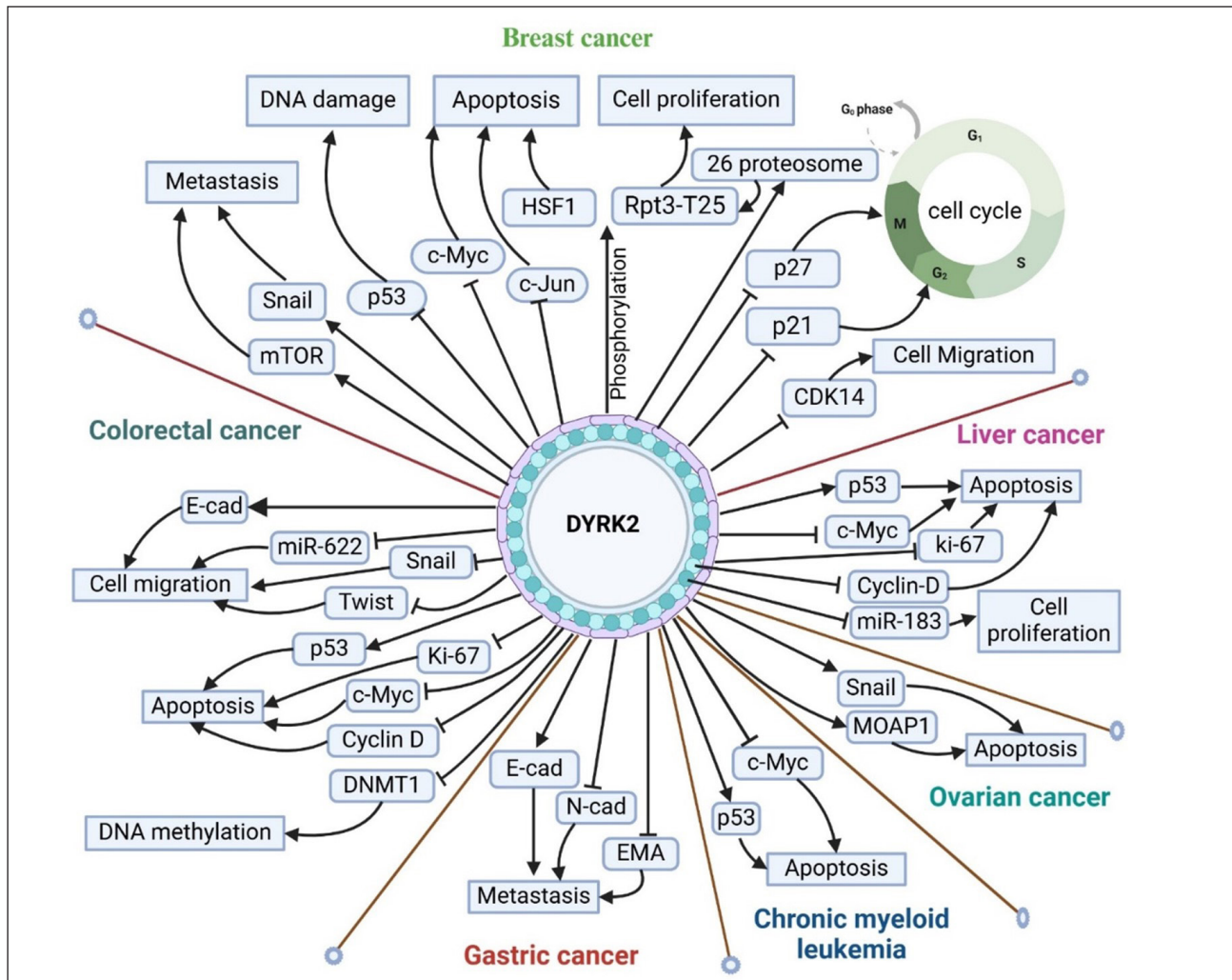


Figure 4: Overview of the roles of DYRK2 in a variety of cancers. HSF1: heat shock factor 1; mTOR: mammalian target of rapamycin; miR622: microRNA 622; E-cad: E-Cadherin; N-cad: neural cadherin; MOAP1: modulator of apoptosis 1; miR183: microRNA 183; CDK14: cyclin dependent kinase 14; Rpt3-T25: regulatory particle 3-T25; ki-67: kiel-67; CycD: Cyclin D; DNMT1: DNA methylation. Image created in BioRender. Moududee, S. (2024) <https://BioRender.com/k39x947>.

receptor-positive breast cancer [94]. As detailed in a previous review article, DYRK2 may induce apoptosis through phosphorylating c-Jun, c-Myc, and HSF1, inhibit cell stemness by suppressing Kruppel-like factor 4 (KLF4) and telomerase reverse transcriptase (TERT) expression via androgen receptor (AR), and reduce cell proliferation via suppression of CDK14 expression, as well as suppress EMT, invasion, and metastasis through phosphorylation of Snail and induction of mTOR degradation [95].

Colorectal cancer is one of the most common malignant diseases and a leading cause of cancer-related death worldwide [96]. The role of DYRK2 in colorectal cancer has been investigated in cell lines, tissue samples, and xenograft mouse models [28]. DYRK2 phosphorylates p53 to regulate apoptotic cell death in response to DNA damage [27, 79]. Previous studies

have shown that DYRK2-mediated phosphorylation initiates the degradation of several proteins via the ubiquitin-proteasome system. Interestingly, DYRK2 functions as a scaffold for the E3 ubiquitin ligase complex containing E3 isolated by the differential display (EDD), DNA damage-binding protein 1 (DDB1), and Vpr binding protein (VPRBP), and DYRK2-EDVP E3 ligase complex. Meanwhile, the E3 ubiquitin ligase complex has a crucial function in regulating normal mitotic progression because overexpression of both DYRK2 and EDD [97, 98] is frequently reported in cancers [2]. Overexpression of DYRK2 suppresses c-Jun, c-Myc, Ki-67, and Cyclin-D expression, thus inhibiting cell growth, cell migration, invasion, and metastasis of colorectal cancer [96, 99]. DYRK2 plays an important role in epithelial-mesenchymal transition (EMT) by degrading Snail and stabilizing E-

cadherin, leading to inhibition of cell migration and invasion [96]. DYRK2 expression is correlated with an EMT transcription factor TWIST, DNA methylation (DNMT1), and a tumor-related gene (miR-622) in colorectal cancer samples and cell lines [100, 101].

The functional roles of DYRK2 in gastric, leukemia, ovarian, liver, and prostate cancer have also been investigated in patient tissue samples, cell lines, and xenograft mouse models [28]. Taken together, these reports suggest that DYRK2 may be a tumor suppressor or oncogene dependent on the cancer type, which can be targeted differentially according to its role in each type of cancer.

DYRK3 & DYRK4

The roles of DYRK3 and DYRK4 in cancer have not been well studied compared to the other DYRKs [11]. A limited number of studies have revealed the important roles of DYRK3 and DYRK4 in tumorigenesis.

Analysis of DYRK3 substrates has led to identifying consensus phosphorylation sequences for DYRK proteins [102]. DYRK3 promotes cell survival through phosphorylation and activation of silent information regulator sirtuin 1 (SIRT1), an NA-

D⁺-dependent protein deacetylase that has potential in various physiological processes including energy metabolism and stress response. Knockdown of endogenous DYRK1A and DYRK3 leads to hypophosphorylation of SIRT1 and increased acetylation levels of p53 to regulate p53-mediated apoptotic response to genotoxic stress [103]. In organisms across the eukaryotic kingdom, DYRK3 is linked to cellular stress responses, ranging from genotoxic stress, irradiation, nutrient starvation, and osmotic stress [29, 104, 105]. DYRK3 regulates the stability of P-granule-like structures and couples stress granule dissolution to the mechanistic target of rapamycin complex 1 (mTORC1) signaling [30] (Figure 5). Moreover, DYRK3 regulates the phase transition of membrane-less organelles in mitosis [106]. It presents a cyclic partitioning mechanism between stress granules and the cytosol via a low-complexity domain in its N terminus and its kinase activity [30]. A recent study has reported that DYRK3 may influence the formation of plasma membrane-associated platforms (PMAPs) by regulating Liprin- α 1's ability to assemble the protein network that promotes cell motility at the edge of the cell, affecting the formation of PMAPs and the formation and turnover of adhesions at the protruding cell edge [107].

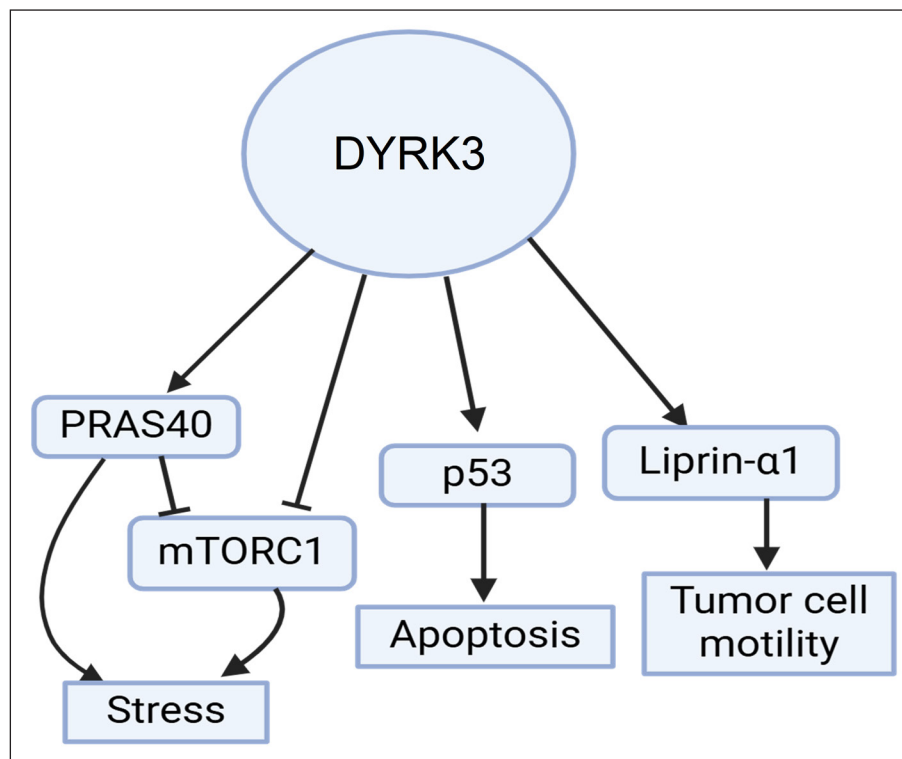


Figure 5: An overview of the interactions between DYRK3 and proteins involved in several biological processes. PRAS40: proline-rich Akt substrate of 40; mTORC1: mammalian target of rapamycin complex 1, Liprin- α 1: Liprin-alpha1. Image created in BioRender. Moududee, S. (2024) <https://BioRender.com/k39x947>.

A few studies have found DYRK3's roles in specific types of tumors. One study has revealed that DYRK3 inhibits hepatocellular carcinoma (HCC) [108], whereas another study has shown that DYRK3 was upregulated in neuroblastoma patients with poor prognosis [109]. DYRK3 may be linked to the aggressiveness of glioblastoma [110, 111], and downregulation of DYRK3 impairs glioblastoma cell migration and invasion [112]. A recent study has shown that DYRK3 is a tumor-promoting oncogene in serous ovarian cancer (SOC) as DYRK3 knockdown inhibits cell viability, invasion, and in vivo tumor growth [113]. Overall, DYRK3's roles in cancer are complicated and cancer-type dependent.

DYRK4 is the fourth member of the DYRK subfamily, which has not been well studied in cancer. One study has found that phosphorylation of the second tyrosine residue within the activation loop appears to have the potential for the full activity of DYRK4 [31]. DYRK4 exhibits different substrate specificity, unlike other members of the DYRK family, with 24 out of 30 peptide substrates containing a serine/threonine residue followed by a proline, indicating that DYRK4 may be characterized as a proline-directed kinase [31]. DNA methylation is one of the most common epigenetic modifications and plays a potential role in carcinogenesis [114]. A recent study has revealed an association between DYRK4 hypomethylation in peripheral blood and an increased risk of lung cancer (LC), which suggests the feasibility of blood-based DNA methylation as a new biomarker for the detection of LC [115]. The mechanism and functional implications underlying this association between DYRK4, and LC need further investigation. Another study has reported that overexpression of a neoplastic chimerical transcript RAD51AP1-DYRK4 is associated with luminal B breast cancers (7–17.5 %), and expression of RAD51AP1-DYRK4 is tumor-specific and enriched in estrogen receptor-positive (ER+) luminal B breast tumors (7–18%) compared to luminal A tumors (3–4%) [116]. However, the pathological role of RAD51AP-DYRK4 in luminal B breast cancer remains unclear. Future studies will be needed to pinpoint the mechanisms engaged by RAD51AP1-DYRK4 to endow breast cancer cells.

Conclusion

Current literature reports indicate that DYRKs are a new class of 'kinase of interest' in human diseases including cancer. However, the roles of DYRKs in

cancer have not been fully elucidated, although there are many findings from studying DYRK expression in cancer cell lines, tumor tissues, and xenografted mouse models. DYRKs have been shown to play tumor-suppressor roles or oncogenic roles, depending on the specific DYRK members and types of cancer. More studies are needed to understand the physiological and pathological functions of DYRK family members in normal and malignant cells. This knowledge will help develop DYRK inhibitors that may have potential in cancer treatment. However, many challenges will be encountered as DYRKs appear to be activated when they are translated, thus lacking an obvious upstream kinase to be targeted. Targeting DYRKs per se or their binding to their substrates may be feasible as shown by a limited number of DYRK inhibitors. Developing a highly selective inhibitor against a specific DYRK member is another challenge, which must be considered as the roles of DYRKs are different.

Acknowledgments

This work was supported in part by a Merit Review Award I01BX004158 from the United States (U.S.) Department of Veterans Affairs, Biomedical Laboratory Research and Development Service. Z.Y. was a Research Physiologist employed by the Research Service, Southeast Louisiana Veterans Health Care System, New Orleans, LA-629. The content of this article is solely the responsibility of the authors and does not necessarily represent the official views or policies of the Department of Veterans Affairs or the United States government. This work was also partially supported by the Tulane Cancer Center and Lavin Bernick Grant.

References

1. Roy PS, BJ: **Cancer and cure A critical analysis.** *Indian J Cancer* 2016, **53**(3):441-442; doi:10.4103/0019-509X.200658.
2. Maddika S, Chen JJ: **Protein kinase DYRK2 is a scaffold that facilitates assembly of an E3 ligase.** *Nat Cell Biol* 2009, **11**(4):409-U110; doi:10.1038/ncb1848.
3. Becker W: **A wake-up call to quiescent cancer cells - potential use of DYRK1B inhibitors in cancer therapy.** *Febs J* 2018, **285**(7):1203-1211; doi:10.1111/febs.14347.

4. Collett MSE, R.L.: **Protein kinase activity associated with the avian sarcoma virus src gene product.** *Proc Natl Acad Sci USA* 1978, **75**(4) doi:10.1073/pnas.75.4.2021.
5. Bhullar KS, Lagaron NO, McGowan EM, Parmar I, Jha A, Hubbard BP, Rupasinghe HPV: **Kinase-targeted cancer therapies: progress, challenges and future directions.** *Mol Cancer* 2018, **17**(1):48; doi:10.1186/s12943-018-0804-2; PMC5817855.
6. Manning G, Whyte DB, Martinez R, Hunter T, Sudarsanam S: **The protein kinase complement of the human genome.** *Science* 2002, **298**(5600):1912-1934; doi:10.1126/science.1075762.
7. Van der Laden J, Soppa U, Becker W: **Effect of tyrosine autophosphorylation on catalytic activity and subcellular localization of homeodomain-interacting protein kinases (HIPK).** *Cell Commun Signal* 2015, **13**doi:ARTN 310.1186/s12964-014-0082-6.
8. Himpel S, Panzer P, Eirmbter K, Czajkowska H, Sayed M, Packman LC, Blundell T, Kentrup H, Grötzinger J, Joost HG *et al.*: **Identification of the autophosphorylation sites and characterization of their effects in the protein kinase DYRK1A.** *Biochem J* 2001, **359**:497-505; doi:DOI 10.1042/0264-6021:3590497.
9. Arron JR, Winslow MM, Polleri A, Chang CP, Wu H, Gao X, Neilson JR, Chen L, Heit JJ, Kim SK *et al.*: **NFAT dysregulation by increased dosage of and on chromosome 21.** *Nature* 2006, **441**(7093):595-600; doi:10.1038/nature04678.
10. Stephen E. Mercer DZE, Xiaobing Deng, Seung-hwan Lim, Thomas R. Mazur, and Eileen Friedman: **Mirk/Dyrk1B Mediates Survival during the Differentiation of C2C12 Myoblasts.** *The Journal of Biological Chemistry* 2005doi:DOI 10.1074/jbc.M413594200.
11. Boni J, Rubio-Perez C, López-Bigas N, Fillat C, de la Luna S: **The DYRK Family of Kinases in Cancer: Molecular Functions and Therapeutic Opportunities.** *Cancers* 2020, **12**(8)doi:ARTN 210610.3390/cancers12082106.
12. Nora Gockler GJ, Chrisovalantis Papadopoulos, Ulf Soppa, Francisco J. Tejedor and Walter Becker.: **Harmine specifically inhibits protein kinase DYRK1A and interferes with neurite formation.** *The FEBS Journal* 2009doi:doi:10.1111/j.1742-4658.2009.07346.x.
13. Lochhead PA, Sibbet G, Morrice N, Cleghon V: **Activation-loop autophosphorylation is mediated by a novel transitional intermediate form of DYRKs.** *Cell* 2005, **121**(6):925-936; doi:10.1016/j.cell.2005.03.034.
14. Walter Becker YW, Kristiane Wetzel, Klaus Eirmbter, Francisco J. Tejedor, and Hans-Georg Joost: **Sequence Characteristics, Subcellular Localization, and Substrate Specificity of DYRK-related Kinases, a Novel Family of Dual Specificity Protein Kinases.** *The Journal of Biological chemistry* 1998, **273**(40):25893-25902; doi:10.1074/jbc.273.40.25893.
15. Alvarez M, Estivill X, de la Luna S: **DYRK1A accumulates in splicing speckles through a novel targeting signal and induces speckle disassembly.** *J Cell Sci* 2003, **116**(15):3099-3107; doi:DOI 10.1242/jcs.00618.
16. Ross Kinstrie NL, Diego Miranda-Saavedra, Gary Sibbet, Jingfen Han, Pamela A. Lochhead, Vaughn Cleghon: **Characterization of a Domain that Transiently Converts Class 2 DYRKs into Intramolecular Tyrosine Kinases.** *Sci Signal* 2010, **3**(111)doi:10.1126/scisignal.2000579.
17. Maria L. Arbones Aurore Thomazeau AN-K, Masatoshi Hagiwara, Jean M. Delabar **DYRK1A and cognition: A lifelong relationship.** *Pharmacology & Therapeutics* 2019, **194**:199-221; doi:10.1016/j.pharmthera.2018.09.010.
18. Friedman E: **Mirk/Dyrk1B in cancer.** *Journal of Cellular Biochemistry* 2007, **102**(2):274-279; doi:10.1002/jcb.21451.
19. Larisa Litovchick LAF, Selene K. Swanson, Michael P. Washburn, and James A. DeCaprio: **DYRK1A protein kinase promotes quiescence and senescence through DREAM complex assembly.** *Genes Dev* 2011, **25**(8):801-813; doi:10.1101/gad.2034211.
20. Vassiliki Fotaki MD, Soledad Alcántara, Salvador Martínez, Eulàlia Martí, Caty Casas, Joana Visa, Eduardo Soriano, Xavier Estivill, Maria L Arbonés: **Dyrk1A Haploinsufficiency Affects Viability and Causes Developmental Delay and Abnormal Brain Morphology in Mice.** *Molecular and cellular Biology* 2002, **22**(18):6636-6647; doi:10.1128/MCB.22.18.6636-6647.
21. Ariadna Laguna SA, María José Barallobre, Susana de la Luna, Pedro de la Villa, Maria L. Arbonés: **The Protein Kinase DYRK1A Regulates Caspase-9-Mediated Apoptosis during Retina Development.** *Developmental Cell* 2008, **15**(6):841-853; doi.
22. Yang EJ, Ahn YS, Chung KC: **Protein kinase Dyrk1 activates cAMP response element-bind-**

- ing protein during neuronal differentiation in hippocampal progenitor cells. *J Biol Chem* 2001, **276**(43):39819-39824; doi:10.1074/jbc.M104091200.
23. **Characterization of a Domain that Transiently Converts Class 2 DYRKs into Intramolecular-Tyrosine Kinases.** doi.
 24. Becker W: **Emerging role of DYRK family protein kinases as regulators of protein stability in cell cycle control.** *Cell Cycle* 2012, **11**(18):3389-3394; doi:10.4161/cc.21404.
 25. Mercer SE, Friedman E: **Mirk/Dyrk1B: a multifunctional dual-specificity kinase involved in growth arrest, differentiation, and cell survival.** *Cell Biochem Biophys* 2006, **45**(3):303-315; doi:10.1385/CBB:45:3:303.
 26. Dong C, West KL, Tan XY, Li JS, Ishibashi T, Yu CH, Sy SMH, Leung JWC, Huen MSY: **Screen identifies DYRK1B network as mediator of transcription repression on damaged chromatin.** *P Natl Acad Sci USA* 2020, **117**(29):17019-17030; doi:10.1073/pnas.2002193117.
 27. Taira N, Nihira K, Yamaguchi T, Miki Y, Yoshida K: **DYRK2 is targeted to the nucleus and controls p53 via Ser46 phosphorylation in the apoptotic response to DNA damage.** *Mol Cell* 2007, **25**(5):725-738; doi:10.1016/j.molcel.2007.02.007.
 28. Yoshida S, Aoki K, Fujiwara K, Nakakura T, Kawamura A, Yamada K, Ono M, Yogosawa S, Yoshida K: **The novel ciliogenesis regulator DYRK2 governs Hedgehog signaling during mouse embryogenesis.** *Elife* 2020, **9**;doi:10.7554/eLife.57381.; PMC7410489.
 29. Zhang D, Li K, Erickson-Miller CL, Weiss M, Wojchowski DM: **DYRK gene structure and erythroid-restricted features of DYRK3 gene expression.** *Genomics* 2005, **85**(1):117-130; doi:10.1016/j.ygeno.2004.08.021.
 30. Wippich F, Bodenmiller B, Trajkovska MG, Wanka S, Aebersold R, Pelkmans L: **Dual Specificity Kinase DYRK3 Couples Stress Granule Condensation/Dissolution to mTORC1 Signaling.** *Cell* 2013, **152**(4):791-805; doi:10.1016/j.cell.2013.01.033.
 31. Papadopoulos C, Arato K, Lilienthal E, Zerweck J, Schutkowski M, Chatain N, Müller-Newen G, Becker W, de la Luna S: **Splice Variants of the Dual Specificity Tyrosine Phosphorylation-regulated Kinase 4 (DYRK4) Differ in Their Subcellular Localization and Catalytic Activity.** *Journal of Biological Chemistry* 2011, **286**(7):5494-5505; doi:10.1074/jbc.M110.157909.
 32. Slepak TI, Salay LD, Lemmon VP, Bixby JL: **Dyrk Kinases Regulate Phosphorylation of Doublecortin, Cytoskeletal Organization, and Neuronal Morphology.** *Cytoskeleton* 2012, **69**(7):514-527; doi:10.1002/cm.21021.
 33. Song WJ, Sternberg LR, KastenSportes C, VanKeuren ML, Chung SH, Slack AC, Miller DE, Glover TW, Chiang PW, Lou LD *et al*: **Isolation of human and murine homologues of the Drosophila minibrain gene: Human homologue maps to 21q22.2 in the down syndrome "critical region".** *Genomics* 1996, **38**(3):331-339; doi:DOI 10.1006/geno.1996.0636.
 34. Hasle H, Clemmensen IH, Mikkelsen M: **Risks of leukaemia and solid tumours in individuals with Down's syndrome.** *Lancet* 2000, **355**(9199):165-169; doi:10.1016/S0140-6736(99)05264-2.
 35. Hill DA, Gridley G, Cnattingius S, Mellekmjaer L, Linet M, Adami HO, Olsen JH, Nyren O, Fraumeni JF: **Mortality and cancer incidence among individuals with Down syndrome.** *Arch Intern Med* 2003, **163**(6):705-711; doi:DOI 10.1001/archinte.163.6.705.
 36. Rammohan M, Harris E, Bhansali RS, Zhao E, Li LS, Crispino JD: **The chromosome 21 kinase DYRK1A: emerging roles in cancer biology and potential as a therapeutic target.** *Oncogene* 2022, **41**(14):2003-2011; doi:10.1038/s41388-022-02245-6.
 37. Baek KH, Zaslavsky A, Lynch RC, Britt C, Okada Y, Siarey RJ, Lensch MW, Park IH, Yoon SS, Minami T *et al*: **Down's syndrome suppression of tumour growth and the role of the calcineurin inhibitor DSCR1.** *Nature* 2009, **459**(7250):1126-1130; doi:10.1038/nature08062.; PMC2724004.
 38. Malinge S, Bliss-Moreau M, Kirsammer G, Diebold L, Chlon T, Gurbuxani S, Crispino JD: **Increased dosage of the chromosome 21 ortholog promotes megakaryoblastic leukemia in a murine model of Down syndrome.** *J Clin Invest* 2012, **122**(3):948-962; doi:10.1172/Jci60455.
 39. Chan KS, Sano S, Kataoka K, Abel E, Carbajal S, Beltran L, Clifford J, Peavey M, Shen J, DiGiovanni J: **Forced expression of a constitutively active form of Stat3 in mouse epidermis enhances malignant progression of skin tumors induced by two-stage carcinogenesis.** *Oncogene* 2008, **27**(8):1087-1094; doi:10.1038/sj.onc.1210726.

40. Johnston PA, Grandis JR: **STAT3 signaling: anticancer strategies and challenges.** *Mol Interv* 2011, **11**(1):18-26; doi:10.1124/mi.11.1.4.; PMC3063716.
41. Kurabayashi N, Nguyen MD, Sanada K: **DYRK1A overexpression enhances STAT activity and astroglioneogenesis in a Down syndrome mouse model.** *EMBO Rep* 2015, **16**(11):1548-1562; doi:10.15252/embr.201540374.; PMC4641506.
42. Siveen KS, Sikka S, Surana R, Dai XY, Zhang JW, Kumar AP, Tan BKH, Sethi G, Bishayee A: **Targeting the STAT3 signaling pathway in cancer: Role of synthetic and natural inhibitors.** *Bba-Rev Cancer* 2014, **1845**(2):136-154; doi:10.1016/j.bbcan.2013.12.005.
43. Li YL, Ding K, Hu X, Wu LW, Zhou DM, Rao MJ, Lin NM, Zhang C: **DYRK1A inhibition suppresses STAT3/EGFR/Met signalling and sensitizes EGFR wild-type NSCLC cells to AZD9291.** *J Cell Mol Med* 2019, **23**(11):7427-7437; doi:10.1111/jcmm.14609.
44. Altaba ARI: **Hedgehog Signaling and the Gli Code in Stem Cells, Cancer, and Metastases.** *Science Signaling* 2011, **4**(200)doi:ARTN pt910.1126/scisignal.2002540.
45. Zhou BH, Lin WL, Long YL, Yang YK, Zhang H, Wu KM, Chu Q: **Notch signaling pathway: architecture, disease, and therapeutics.** *Signal Transduct Tar* 2022, **7**(1)doi:ARTN 9510.1038/s41392-022-00934-y.
46. Lobry C, Oh P, Mansour MR, Look AT, Aifantis I: **Notch signaling: switching an oncogene to a tumor suppressor.** *Blood* 2014, **123**(16):2451-2459; doi:10.1182/blood-2013-08-355818.
47. Lawinger P, Venugopal R, Guo ZS, Immaneni A, Sengupta D, Lu WY, Rastelli L, Carneiro AMD, Levin V, Fuller GN *et al*: **The neuronal repressor REST/NRSF is an essential regulator in medulloblastoma cells.** *Nat Med* 2000, **6**(7):826-831; doi:10.1038/77565.
48. Su XH, Gopalakrishnan V, Stearns D, Aldape K, Lang FF, Fuller G, Snyder E, Eberhart CG, Majumder S: **Abnormal expression of REST/NRSF and Myc in neural stem/progenitor cells causes cerebellar tumors by blocking neuronal differentiation.** *Molecular and Cellular Biology* 2006, **26**(5):1666-1678; doi:10.1128/Mcb.26.5.1666-1678.2006.
49. Palm K, Metsis M, Timmusk T: **Neuron-specific splicing of zinc finger transcription factor REST/NRSF/XBR is frequent in neuroblastomas and conserved in human, mouse and rat.** *Mol Brain Res* 1999, **72**(1):30-39; doi:10.1016/S0169-328x(99)00196-5.
50. Kamal MM, Sathyan P, Singh SK, Zinn PO, Marisetty AL, Liang SD, Gumin J, El-Mesallamy HO, Suki D, Colman H *et al*: **REST Regulates Oncogenic Properties of Glioblastoma Stem Cells (vol 30, pg 405, 2012).** *Stem Cells* 2012, **30**(5):1049-1049; doi:10.1002/stem.1085.
51. Fernandez-Martinez J, Vela EM, Tora-Ponsoen M, Ocana OH, Nieto MA, Galceran J: **Attenuation of Notch signalling by the Down-syndrome-associated kinase DYRK1A.** *J Cell Sci* 2009, **122**(10):1574-1583; doi:10.1242/jcs.044354.
52. Gao JC, Yang XJ, Yin P, Hu WF, Liao HF, Miao ZH, Pan C, Li N: **The involvement of FoxO in cell survival and chemosensitivity mediated by Mirk/Dyrk1B in ovarian cancer.** *Int J Oncol* 2012, **40**(4):1203-1209; doi:10.3892/ijo.2011.1293.
53. Gao J, Zheng Z, Rawal B, Schell MJ, Bepler G, Haura EB: **Mirk/Dyrk1B, a novel therapeutic target, mediates cell survival in non-small cell lung cancer cells.** *Cancer Biol Ther* 2009, **8**(17):1671-1679; doi:10.4161/cbt.8.17.9322.; PMC3839311.
54. Chen YY, Wang S, He ZX, Sun FL, Huang YQ, Ni QC, Wang H, Wang YY, Cheng C: **Dyrk1B overexpression is associated with breast cancer growth and a poor prognosis.** *Hum Pathol* 2017, **66**:48-58; doi:10.1016/j.humpath.2017.02.033.
55. Lee K, Deng XB, Friedman E: **Mirk protein kinase is a mitogen-activated protein kinase substrate that mediates survival of colon cancer cells.** *Cancer Res* 2000, **60**(13):3631-3637; doi.
56. Jin K, Ewton DZ, Park S, Hu J, Friedman E: **Mirk Regulates the Exit of Colon Cancer Cells from Quiescence.** *Journal of Biological Chemistry* 2009, **284**(34):22916-22925; doi:10.1074/jbc.M109.035519.
57. Deng XB, Ewton DZ, Li SN, Naqvi A, Mercer SE, Landas S, Friedman E: **The kinase Mirk/Dyrk1B mediates cell survival in pancreatic ductal adenocarcinoma.** *Cancer Res* 2006, **66**(8):4149-4158; doi:10.1158/0008-5472.Can-05-3089.
58. Deng X, Friedman E: **Mirk kinase inhibition blocks the in vivo growth of pancreatic cancer cells.** *Genes Cancer* 2014, **5**(9-10):337-347; doi:10.18632/genesandcancer.29.; PMC4209603.
59. Friedman E: **Mirk/dyrk1B Kinase in Ovarian Cancer.** *Int J Mol Sci* 2013, **14**(3):5560-5575; doi:10.3390/ijms14035560.; PMC3634458.

60. MacKeigan JP, Murphy LO, Blenis J: **Sensitized RNAi screen of human kinases and phosphatases identifies new regulators of apoptosis and chemoresistance.** *Nat Cell Biol* 2005, 7(6):591-U519; doi:10.1038/ncb1258.
61. Tsioras K, Papastefanaki F, Politis PK, Matsas R, Gaitanou M: **Functional Interactions between BM88/Cend1, Ran-Binding Protein M and Dyrk1B Kinase Affect Cyclin D1 Levels and Cell Cycle Progression/Exit in Mouse Neuroblastoma Cells.** *Plos One* 2013, 8(11):ARTN e8217210.1371/journal.pone.0082172.
62. Gaitanou M, Segklia K, Matsas R: **Cend1, a Story with Many Tales: From Regulation of Cell Cycle Progression/Exit of Neural Stem Cells to Brain Structure and Function.** *Stem Cells Int* 2019, 2019:doi:ArtN 205478310.1155/2019/2054783.
63. Zou YL, Ewton DZ, Deng XB, Mercer SE, Friedman E: **Mirk/dyrk1B kinase destabilizes cyclin D1 by phosphorylation at threonine 288.** *Journal of Biological Chemistry* 2004, 279(26):27790-27798; doi:10.1074/jbc.M403042200.
64. Ewton DZ, Hu J, Vilenchik M, Deng XB, Luk KC, Polonskaia A, Hoffman AF, Zipf K, Boylan JF, Friedman EA: **Inactivation of Mirk/Dyrk1b Kinase Targets Quiescent Pancreatic Cancer Cells.** *Mol Cancer Ther* 2011, 10(11):2104-2114; doi:10.1158/1535-7163.Mct-11-0498.
65. Hu J, Nakhla H, Friedman E: **Transient arrest in a quiescent state allows ovarian cancer cells to survive suboptimal growth conditions and is mediated by both Mirk/dyrk1b and p130/RB2.** *Int J Cancer* 2011, 129(2):307-318; doi:10.1002/ijc.25692.
66. Hu J, Friedman E: **Depleting Mirk Kinase Increases Cisplatin Toxicity in Ovarian Cancer Cells.** *Genes Cancer* 2010, 1(8):803-811; doi:10.1177/1947601910377644.; PMC2989622.
67. Hu J, Deng H, Friedman EA: **Ovarian cancer cells, not normal cells, are damaged by Mirk/Dyrk1B kinase inhibition.** *Int J Cancer* 2013, 132(10):2258-2269; doi:10.1002/ijc.27917.; PMC3586305.
68. Li L, Liu Y, Zhang Q, Zhou H, Zhang Y, Yan B: **Comparison of cancer cell survival triggered by microtubule damage after turning Dyrk1B kinase on and off.** *ACS Chem Biol* 2014, 9(3):731-742; doi:10.1021/cb4005589.
69. Gao J, Zhao Y, Lv Y, Chen Y, Wei B, Tian J, Yang Z, Kong F, Pang J, Liu J *et al*: **Mirk/Dyrk1B mediates G0/G1 to S phase cell cycle progression and cell survival involving MAPK/ERK signaling in human cancer cells.** *Cancer Cell Int* 2013, 13(1):2; doi:10.1186/1475-2867-13-2.; PMC3575355.
70. Deng XB, Hu J, Ewton DZ, Friedman E: **Mirk/dyrk1B kinase is upregulated following inhibition of mTOR.** *Carcinogenesis* 2014, 35(9):1968-1976; doi:10.1093/carcin/bgu058.
71. Gruber W, Hutzinger M, Elmer DP, Parigger T, Sternberg C, Cegielski L, Zaja M, Leban J, Michel S, Hamm S *et al*: **DYRK1B as therapeutic target in Hedgehog/GLI-dependent cancer cells with Smoothened inhibitor resistance.** *Oncotarget* 2016, 7(6):7134-7148; doi:10.18632/oncotarget.6910.
72. Song LN, Silva J, Koller A, Rosenthal A, Chen EI, Gelmann EP: **The Tumor Suppressor NKX3.1 Is Targeted for Degradation by DYRK1B Kinase.** *Mol Cancer Res* 2015, 13(5):913-922; doi:10.1158/1541-7786.Mcr-14-0680.
73. Jin K, Park S, Ewton DZ, Friedman E: **The survival kinase Mirk/Dyrk1B is a downstream effector of oncogenic K-ras in pancreatic cancer.** *Cancer Res* 2007, 67(15):7247-7255; doi:10.1158/0008-5472.can-06-4099.
74. Guo X, Wang XR, Wang ZP, Banerjee S, Yang J, Huang L, Dixon JE: **Site-specific proteasome phosphorylation controls cell proliferation and tumorigenesis.** *Nat Cell Biol* 2016, 18(2):202-+; doi:10.1038/ncb3289.
75. Aranda S, Laguna A, de la Luna S: **DYRK family of protein kinases: evolutionary relationships, biochemical properties, and functional roles.** *Faseb J* 2011, 25(2):449-462; doi:10.1096/fj.10-165837.
76. Correa-Sáez A, Jiménez-Izquierdo R, Garrido-Rodríguez M, Morrugares R, Muñoz E, Calzado MA: **Updating dual-specificity tyrosine-phosphorylation-regulated kinase 2 (DYRK2): molecular basis, functions and role in diseases.** *Cell Mol Life Sci* 2020, 77(23):4747-4763; doi:10.1007/s00018-020-03556-1.
77. Lara-Chica M, Correa-Sáez A, Jiménez-Izquierdo R, Garrido-Rodríguez M, Ponce FJ, Moreno R, Morrison K, Di Vona C, Arató K, Jiménez-Jiménez C *et al*: **A novel CDC25A/DYRK2 regulatory switch modulates cell cycle and survival.** *Cell Death Differ* 2022, 29(1):105-117; doi:10.1038/s41418-021-00845-5.

78. Tandon V, de la Vega L, Banerjee S: **Emerging roles of DYRK2 in cancer.** *Journal of Biological Chemistry* 2021, **296**doi:ARTN 10023310.1074/jbc.REV120.015217.
79. Yoshida K: **Nuclear trafficking of pro-apoptotic kinases in response to DNA damage.** *Trends Mol Med* 2008, **14**(7):305-313; doi:10.1016/j.molmed.2008.05.003.
80. Imawari Y, Mimoto R, Hirooka S, Morikawa T, Takeyama H, Yoshida K: **Downregulation of dual-specificity tyrosine-regulated kinase 2 promotes tumor cell proliferation and invasion by enhancing cyclin-dependent kinase 14 expression in breast cancer.** *Cancer Sci* 2017, **109**(2):363-372; doi:10.1111/cas.13459.
81. Gu X, Wang Y, Wang H, Ni Q, Zhang C, Zhu J, Huang W, Xu P, Mao G, Yang S: **Upregulated PFTK1 promotes tumor cell proliferation, migration, and invasion in breast cancer.** *Med Oncol* 2015, **32**(7):195; doi:10.1007/s12032-015-0641-8.
82. Yang L, Zhu J, Huang H, Yang Q, Cai J, Wang Q, Zhu J, Shao M, Xiao J, Cao J *et al*: **PFTK1 Promotes Gastric Cancer Progression by Regulating Proliferation, Migration and Invasion.** *Plos One* 2015, **10**(10):e0140451; doi:10.1371/journal.pone.0140451; PMC4619205.
83. Zhang W, Liu R, Tang C, Xi Q, Lu S, Chen W, Zhu L, Cheng J, Chen Y, Wang W *et al*: **PFTK1 regulates cell proliferation, migration and invasion in epithelial ovarian cancer.** *Int J Biol Macromol* 2016, **85**:405-416; doi:10.1016/j.ijbiomac.2016.01.009.
84. Miyagaki H, Yamasaki M, Miyata H, Takahashi T, Kurokawa Y, Nakajima K, Takiguchi S, Fujiwara Y, Ishii H, Tanaka F *et al*: **Overexpression of PFTK1 predicts resistance to chemotherapy in patients with oesophageal squamous cell carcinoma.** *Br J Cancer* 2012, **106**(5):947-954; doi:10.1038/bjc.2012.35; PMC3305960.
85. Banerjee S, Ji C, Mayfield JE, Goel A, Xiao J, Dixon JE, Guo X: **Ancient drug curcumin impedes 26S proteasome activity by direct inhibition of dual-specificity tyrosine-regulated kinase 2.** *Proc Natl Acad Sci U S A* 2018, **115**(32):8155-8160; doi:10.1073/pnas.1806797115; PMC6094102.
86. Banerjee S, Wei T, Wang J, Lee JJ, Gutierrez HL, Chapman O, Wiley SE, Mayfield JE, Tandon V, Juarez EF *et al*: **Inhibition of dual-specificity tyrosine phosphorylation-regulated kinase 2 perturbs 26S proteasome-addicted neoplastic progression.** *Proc Natl Acad Sci U S A* 2019, **116**(49):24881-24891; doi:10.1073/pnas.1912033116; PMC6900511.
87. Taira N, Yamamoto H, Yamaguchi T, Miki Y, Yoshida K: **ATM augments nuclear stabilization of DYRK2 by inhibiting MDM2 in the apoptotic response to DNA damage.** *J Biol Chem* 2010, **285**(7):4909-4919; doi:10.1074/jbc.M109.042341; PMC2836095.
88. Taira N, Mimoto R, Kurata M, Yamaguchi T, Kitagawa M, Miki Y, Yoshida K: **DYRK2 priming phosphorylation of c-Jun and c-Myc modulates cell cycle progression in human cancer cells.** *J Clin Invest* 2012, **122**(3):859-872; doi:10.1172/JCI60818; PMC3287383.
89. Pérez M, García-Limones C, Zapico I, Marina A, Schmitz ML, Muñoz E, Calzado MA: **Mutual regulation between SIAH2 and DYRK2 controls hypoxic and genotoxic signaling pathways.** *J Mol Cell Biol* 2012, **4**(5):316-330; doi:10.1093/jmcb/mjs047.
90. Morrugares R, Correa-Saez A, Moreno R, Garrido-Rodriguez M, Munoz E, de la Vega L, Calzado MA: **Phosphorylation-dependent regulation of the NOTCH1 intracellular domain by dual-specificity tyrosine-regulated kinase 2.** *Cell Mol Life Sci* 2020, **77**(13):2621-2639; doi:10.1007/s00018-019-03309-9; PMC7320039.
91. Donnelly N, Storchova Z: **Causes and consequences of protein folding stress in aneuploid cells.** *Cell Cycle* 2015, **14**(4):495-501; doi:10.1080/15384101.2015.1006043; PMC4347676.
92. Ohashi A, Ohori M, Iwai K, Nakayama Y, Nambu T, Morishita D, Kawamoto T, Miyamoto M, Hirayama T, Okaniwa M *et al*: **Aneuploidy generates proteotoxic stress and DNA damage concurrently with p53-mediated post-mitotic apoptosis in SAC-impaired cells.** *Nat Commun* 2015, **6**:7668; doi:10.1038/ncomms8668; PMC4506520.
93. Mimoto R, Taira N, Takahashi H, Yamaguchi T, Okabe M, Uchida K, Miki Y, Yoshida K: **DYRK2 controls the epithelial-mesenchymal transition in breast cancer by degrading Snail.** *Cancer Lett* 2013, **339**(2):214-225; doi:10.1016/j.canlet.2013.06.005.
94. Mimoto R, Nihira NT, Hirooka S, Takeyama H, Yoshida K: **Diminished DYRK2 sensitizes hormone receptor-positive breast cancer to ever-**

- olimus by the escape from degrading mTOR.** *Cancer Lett* 2017, **384**:27-38; doi:10.1016/j.canlet.2016.10.015.
95. Mochimaru Y, Yoshida K: **Functional Roles of DYRK2 as a Tumor Regulator.** *Curr Issues Mol Biol* 2023, **45**(10):8539-8551; doi:10.3390/cimb45100538.
 96. Ito D, Yogosawa S, Mimoto R, Hirooka S, Horiuchi T, Eto K, Yanaga K, Yoshida K: **Dual-specificity tyrosine-regulated kinase 2 is a suppressor and potential prognostic marker for liver metastasis of colorectal cancer.** *Cancer Sci* 2017, **108**(8):1565-1573; doi:10.1111/cas.13280.
 97. Clancy JL, Henderson MJ, Russell AJ, Anderson DW, Bova RJ, Campbell IG, Choong DYH, Macdonald GA, Mann GJ, Nolan T *et al*: **EDD, the human orthologue of the tumour suppressor gene, is amplified and overexpressed in cancer.** *Oncogene* 2003, **22**(32):5070-5081; doi:10.1038/sj.onc.1206775.
 98. O'Brien PM, Davies MJ, Scurry JP, Smith AN, Barton CA, Henderson MJ, Saunders DN, Gloss BS, Patterson KI, Clancy JL *et al*: **The E3 ubiquitin ligase EDD is an adverse prognostic factor for serous epithelial ovarian cancer and modulates cisplatin resistance (vol 98, pg 1085, 2008).** *Brit J Cancer* 2008, **98**(11):1880-1880; doi:10.1038/sj.bjc.6604419.
 99. Imaizumi Y, Yoshida S, Kanegae Y, Eto K, Yoshida K: **Enforced dual-specificity tyrosine-regulated kinase 2 expression by adenovirus-mediated gene transfer inhibits tumor growth and metastasis of colorectal cancer.** *Cancer Sci* 2022, **113**(3):960-970; doi:10.1111/cas.15247; PMC8898707.
 100. Wang Y, Sun J, Wei XL, Luan L, Zeng XD, Wang CF, Zhao W: **Decrease of miR-622 expression suppresses migration and invasion by targeting regulation of DYRK2 in colorectal cancer cells.** *Oncotargets Ther* 2017, **10**:1091-1100; doi:10.2147/Ott.S125724.
 101. Wu C, Sun G, Wang F, Chen J, Zhan F, Lian X, Wang J, Weng F, Li B, Tang W *et al*: **DYRK2 downregulation in colorectal cancer leads to epithelial-mesenchymal transition induction and chemoresistance.** *Sci Rep* 2022, **12**(1):22496; doi:10.1038/s41598-022-25053-0; PMC9797492.
 102. Campbell LE, Proud CG: **Differing substrate specificities of members of the DYRK family of arginine-directed protein kinases.** *FEBS Lett* 2002, **510**(1-2):31-36; doi:10.1016/s0014-5793(01)03221-5.
 103. Guo X, Williams JG, Schug TT, Li X: **DYRK1A and DYRK3 promote cell survival through phosphorylation and activation of SIRT1.** *J Biol Chem* 2010, **285**(17):13223-13232; doi:10.1074/jbc.M110.102574; PMC2857074.
 104. Taminato A, Bagattini R, Gorjao R, Chen GK, Kuspa A, Souza GM: **Role for YAP, cAMP, and protein kinase A in regulation of stress responses of cells.** *Mol Biol Cell* 2002, **13**(7):2266-2275; doi:10.1091/mbc.01-11-0555.
 105. Seifert A, Clarke PR: **p38 α - and DYRK1A-dependent phosphorylation of caspase-9 at an inhibitory site in response to hyperosmotic stress.** *Cell Signal* 2009, **21**(11):1626-1633; doi:10.1016/j.cellsig.2009.06.009.
 106. Rai AK, Chen JX, Selbach M, Pelkmans L: **Kinase-controlled phase transition of membraneless organelles in mitosis.** *Nature* 2018, **559**(7713):211-216; doi:10.1038/s41586-018-0279-8.
 107. Ramella M, Ribolla LM, Surini S, Sala K, Tonoli D, Cioni JM, Rai AK, Pelkmans L, de Curtis I: **Dual specificity kinase DYRK3 regulates cell migration by influencing the stability of protrusions.** *iScience* 2024, **27**(4):109440; doi:10.1016/j.isci.2024.109440; PMC10952033.
 108. Ma F, Zhu Y, Liu X, Zhou Q, Hong X, Qu C, Feng X, Zhang Y, Ding Q, Zhao J *et al*: **Dual-Specificity Tyrosine Phosphorylation-Regulated Kinase 3 Loss Activates Purine Metabolism and Promotes Hepatocellular Carcinoma Progression.** *Hepatology* 2019, **70**(5):1785-1803; doi:10.1002/hep.30703.
 109. Uhl KL, Schultz CR, Geerts D, Bachmann AS: **Harmine, a dual-specificity tyrosine phosphorylation-regulated kinase (DYRK) inhibitor induces caspase-mediated apoptosis in neuroblastoma.** *Cancer Cell Int* 2018, **18**:82; doi:10.1186/s12935-018-0574-3; PMC5992763.
 110. Kim K, Lee S, Kang H, Shin E, Kim HY, Youn H, Youn B: **Dual Specificity Kinase DYRK3 Promotes Aggressiveness of Glioblastoma by Altering Mitochondrial Morphology and Function.** *Int J Mol Sci* 2021, **22**(6):doi:10.3390/ijms22062982; PMC8000785.
 111. Kang G, Hwang WC, Do IG, Wang K, Kang SY, Lee J, Park SH, Park JO, Kang WK, Jang J *et al*: **Exome Sequencing Identifies Early Gas-**

- tric Carcinoma as an Early Stage of Advanced Gastric Cancer.** *Plos One* 2013, **8**(12)doi:ARTN e8277010.1371/journal.pone.0082770.
112. Ivanova E, Sharma SD, Brichkina A, Pfefferle P, Keber U, Pagenstecher A, Lauth M: **DYRK3 contributes to differentiation and hypoxic control in neuroblastoma.** *Biochem Biophys Res Commun* 2021, **567**:215-221; doi:10.1016/j.bbrc.2021.06.053.
 113. Sun J, Zhang Y, Li A, Yu H: **Dual-Specificity Tyrosine Phosphorylation-Regulated Kinase 3 Expression and Its Correlation with Prognosis and Growth of Serous Ovarian Cancer: Correlation of DYRK3 with Ovarian Cancer Survival.** *Int J Genomics* 2024, **2024**:6683202; doi:10.1155/2024/6683202.; PMC10963101.
 114. Robertson KD: **DNA methylation and human disease.** *Nature Reviews Genetics* 2005, **6**:597–610; doi.
 115. Qiao R, Zhu Q, Di FF, Liu CL, Song YK, Zhang J, Xu T, Wang Y, Dai LP, Gu WJ *et al*: **Hypomethylation of in peripheral blood is associated with increased lung cancer risk.** *Mol Carcinogen* 2023, **62**(11):1745-1754; doi:10.1002/mc.23612.
 116. Liu CC, Veeraraghavan J, Tan Y, Kim JA, Wang X, Loo SK, Lee S, Hu Y, Wang XS: **A Novel Neoplastic Fusion Transcript, RAD51AP1-DYRK4, Confers Sensitivity to the MEK Inhibitor Trametinib in Aggressive Breast Cancers.** *Clin Cancer Res* 2021, **27**(3):785-798; doi:10.1158/1078-0432.CCR-20-2769.; PMC7934498.